

JOINT INVENTORS

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Richard Zimmermann

APPLICATION FOR UNITED STATES LETTERS PATENT

S P E C I F I C A T I O N

TO ALL WHOM IT MAY CONCERN:

Be it known that we, Timothy J. Martins, a citizen of the United States of America, residing at 3115 209th Street SE, Bothell (98021), in the County of King and State of Washington; and Kerry W. Fowler, a citizen of the United States of America, residing at 747 North 66th Street, Seattle (98103), in the County of King and State of Washington; and Joshua Odingo, a citizen of Kenya, residing at 20415 Bothell-Everett Hwy. #G201, Bothell (98021), in the County of King and State of Washington; and Laurence E. Burgess, a citizen of The United States of America, residing at 5562 High Country Court, Boulder (80301), in the County of Boulder and

CYCLIC AMP-SPECIFIC PHOSPHODIESTERASE INHIBITORS

CROSS-REFERENCE TO RELATED APPLICATION

5 This application claims benefit of provisional application Serial No. 60/172,023, filed December 23, 1999.

FIELD OF INVENTION

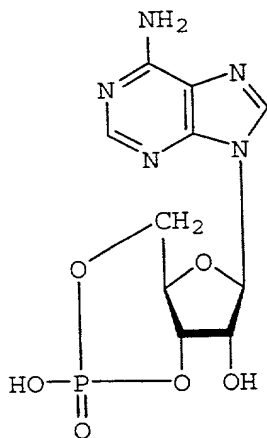
10 The present invention relates to a series of compounds that are potent and selective inhibitors of cyclic adenosine 3',5'-monophosphate specific phosphodiesterase (cAMP specific PDE). In
15 particular, the present invention relates to a series of novel compounds that are useful for inhibiting the function of cAMP specific PDE, in particular, PDE4, as well as methods of making the same, pharmaceutical compositions containing the same, and
20 their use as therapeutic agents, for example, in treating inflammatory diseases and other diseases involving elevated levels of cytokines and proinflammatory mediators.

25 BACKGROUND OF THE INVENTION

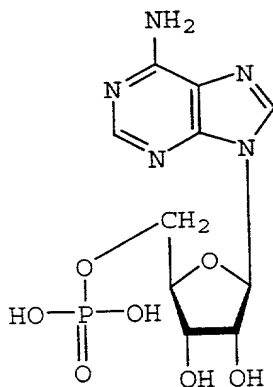
 Chronic inflammation is a multi-factorial disease complication characterized by activation of multiple types of inflammatory cells, particularly
30 cells of lymphoid lineage (including T lymphocytes) and myeloid lineage (including granulocytes, macrophages, and monocytes). Proinflammatory mediators, including cytokines, such as tumor necrosis factor

(TNF) and interleukin-1 (IL-1), are produced by these activated cells. Accordingly, an agent that suppresses the activation of these cells, or their production of proinflammatory cytokines, would be
5 useful in the therapeutic treatment of inflammatory diseases and other diseases involving elevated levels of cytokines.

Cyclic adenosine monophosphate (cAMP) is a second messenger that mediates the biologic re-
10 sponses of cells to a wide range of extracellular stimuli. When the appropriate agonist binds to specific cell surface receptors, adenylate cyclase is activated to convert adenosine triphosphate (ATP) to cAMP. It is theorized that the agonist induced
15 actions of cAMP within the cell are mediated predominantly by the action of cAMP-dependent protein kinases. The intracellular actions of cAMP are terminated by either a transport of the nucleotide to the outside of the cell, or by enzymatic cleavage
20 by cyclic nucleotide phosphodiesterases (PDEs), which hydrolyze the 3'-phosphodiester bond to form 5'-adenosine monophosphate (5'-AMP). 5'-AMP is an inactive metabolite. The structures of cAMP and 5'-AMP are illustrated below.



cAMP



5'-AMP

Elevated levels of cAMP in human myeloid and lymphoid lineage cells are associated with the suppression of cell activation. The intracellular enzyme family of PDEs, therefore, regulates the level of cAMP in cells. PDE4 is a predominant PDE isotype in these cells, and is a major contributor to cAMP degradation. Accordingly, the inhibition of

PDE function would prevent the conversion of cAMP to the inactive metabolite 5'-AMP and, consequently, maintain higher cAMP levels, and, accordingly, suppress cell activation (see Beavo et al., "Cyclic Nucleotide Phosphodiesterases: Structure, Regulation and Drug Action," Wiley and Sons, Chichester, pp. 3-14, (1990)); Torphy et al., *Drug News and Perspectives*, 6, pp. 203-214 (1993); Giembycz et al., *Clin. Exp. Allergy*, 22, pp. 337-344 (1992)).

- 10 In particular, PDE4 inhibitors, such as rolipram, have been shown to inhibit production of TNF α and partially inhibit IL-1 β release by monocytes (see Semmler et al., *Int. J. Immunopharmacol.*, 15, pp. 409-413 (1993); Molnar-Kimber et al., *Mediators of Inflammation*, 1, pp. 411-417 (1992)). PDE4 inhibitors also have been shown to inhibit the production of superoxide radicals from human polymorphonuclear leukocytes (see Verghese et al., *J. Mol. Cell. Cardiol.*, 21 (Suppl. 2), S61 (1989);
- 15 Nielson et al., *J. Allergy Immunol.*, 86, pp. 801-808 (1990)); to inhibit the release of vasoactive amines and prostanoids from human basophils (see Peachell et al., *J. Immunol.*, 148, pp. 2503-2510 (1992)); to inhibit respiratory bursts in eosinophils (see Dent et al., *J. Pharmacol.*, 103, pp. 1339-1346 (1991));
- 25 and to inhibit the activation of human T-lymphocytes (see Robicsek et al., *Biochem. Pharmacol.*, 42, pp. 869-877 (1991)).

- Inflammatory cell activation and excessive
- 30 or unregulated cytokine (e.g., TNF α and IL-1 β) production are implicated in allergic, autoimmune, and inflammatory diseases and disorders, such as rheuma-

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toid arthritis, osteoarthritis, gouty arthritis,
spondylitis, thyroid associated ophthalmopathy,
Behcet's disease, sepsis, septic shock, endotoxic
shock, gram negative sepsis, gram positive sepsis,
5 toxic shock syndrome, asthma, chronic bronchitis,
adult respiratory distress syndrome, chronic pulmo-
nary inflammatory disease, such as chronic obstruc-
tive pulmonary disease, silicosis, pulmonary sarco-
idosis, reperfusion injury of the myocardium, brain,
10 and extremities, fibrosis, cystic fibrosis, keloid
formation, scar formation, atherosclerosis, trans-
plant rejection disorders, such as graft vs. host
reaction and allograft rejection, chronic glomerulo-
nephritis, lupus, inflammatory bowel disease, such
15 as Crohn's disease and ulcerative colitis, prolifer-
ative lymphocyte diseases, such as leukemia, and
inflammatory dermatoses, such as atopic dermatitis,
psoriasis, and urticaria.

Other conditions characterized by elevated
20 cytokine levels include brain injury due to moderate
trauma (see Dhillon et al., *J. Neurotrauma*, 12, pp.
1035-1043 (1995); Suttorp et al., *J. Clin. Invest.*,
91, pp. 1421-1428 (1993)), cardiomyopathies, such as
congestive heart failure (see Bristow et al., *Circu-*
25 *lation*, 97, pp. 1340-1341 (1998)), cachexia, cachex-
ia secondary to infection or malignancy, cachexia
secondary to acquired immune deficiency syndrome
(AIDS), ARC (AIDS related complex), fever myalgias
due to infection, cerebral malaria, osteoporosis and
30 bone resorption diseases, keloid formation, scar
tissue formation, and pyrexia.

In particular, TNF α has been identified as
having a role with respect to human acquired immune

deficiency syndrome (AIDS). AIDS results from the infection of T-lymphocytes with Human Immunodeficiency Virus (HIV). Although HIV also infects and is maintained in myeloid lineage cells, TNF has been shown to upregulate HIV infection in T-lymphocytic and monocytic cells (see Poli et al., *Proc. Natl. Acad. Sci. USA*, 87, pp. 782-785 (1990)).

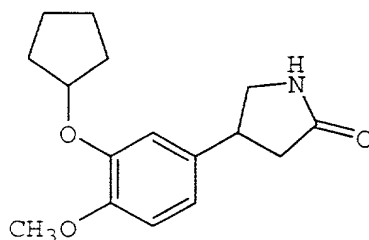
Several properties of TNF α , such as stimulation of collagenases, stimulation of angiogenesis *in vivo*, stimulation of bone resorption, and an ability to increase the adherence of tumor cells to endothelium, are consistent with a role for TNF in the development and metastatic spread of cancer in the host. TNF α recently has been directly implicated in the promotion of growth and metastasis of tumor cells (see Orosz et al., *J. Exp. Med.*, 177, pp. 1391-1398 (1993)).

PDE4 has a wide tissue distribution. There are at least four genes for PDE4 of which multiple transcripts from any given gene can yield several different proteins that share identical catalytic sites. The amino acid identity between the four possible catalytic sites is greater than 85%. Their shared sensitivity to inhibitors and their kinetic similarity reflect the functional aspect of this level of amino acid identity. It is theorized that the role of these alternatively expressed PDE4 proteins allows a mechanism by which a cell can differentially localize these enzymes intracellularly and/or regulate the catalytic efficiency via post translational modification. Any given cell type that expresses the PDE4 enzyme typi-

cally expresses more than one of the four possible genes encoding these proteins.

Investigators have shown considerable interest in the use of PDE4 inhibitors as anti-inflammatory agents. Early evidence indicates that PDE4 inhibition has beneficial effects on a variety of inflammatory cells such as monocytes, macrophages, T-cells of the Th-1 lineage, and granulocytes. The synthesis and/or release of many proinflammatory mediators, such as cytokines, lipid mediators, superoxide, and biogenic amines, such as histamine, have been attenuated in these cells by the action of PDE4 inhibitors. The PDE4 inhibitors also affect other cellular functions including T-cell proliferation, granulocyte transmigration in response to chemotoxic substances, and integrity of endothelial cell junctions within the vasculature.

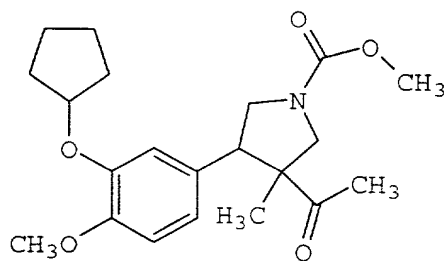
The design, synthesis, and screening of various PDE4 inhibitors have been reported. Methylxanthines, such as caffeine and theophylline, were the first PDE inhibitors discovered, but these compounds are nonselective with respect to which PDE is inhibited. The drug rolipram, an antidepressant agent, was one of the first reported specific PDE4 inhibitors. Rolipram, having the following structural formula, has a reported 50% Inhibitory Concentration (IC_{50}) of about 200 nM (nanomolar) with respect to inhibiting recombinant human PDE4.



Rolipram

Investigators have continued to search for PDE4 inhibitors that are more selective with respect to inhibiting PDE4, that have a lower IC₅₀ than rolipram, and that avoid the undesirable central nervous system (CNS) side effects, such as retching, vomiting, and sedation, associated with the administration of rolipram. One class of compounds is disclosed in Feldman et al. U.S. Patent No. 5,665,754. The compounds disclosed therein are substituted pyrrolidines having a structure similar to rolipram. One particular compound, having structural formula (I), has an IC₅₀ with respect to human recombinant PDE4 of about 2 nM. Inasmuch as a favorable separation of emetic side effect from efficacy was observed, these compounds did not exhibit a reduction in undesirable CNS effects.

5



(I)

10 In addition, several companies are now
undertaking clinical trials of other PDE4 inhibi-
tors. However, problems relating to efficacy and
adverse side effects, such as emesis and central
nervous system disturbances, remain unsolved.

15 Accordingly, compounds that selectively
inhibit PDE4, and that reduce or eliminate the ad-
verse CNS side effects associated with prior PDE4
inhibitors, would be useful in the treatment of
allergic and inflammatory diseases, and other dis-
20 eases associated with excessive or unregulated pro-
duction of cytokines, such as TNF. In addition,
selective PDE4 inhibitors would be useful in the
treatment of diseases that are associated with ele-
vated cAMP levels or PDE4 function in a particular
25 target tissue.

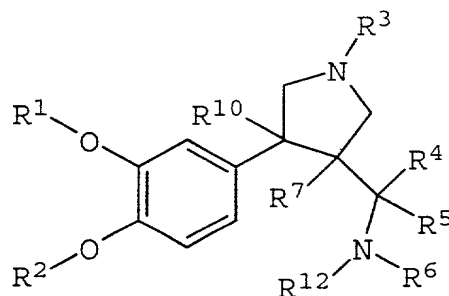
SUMMARY OF THE INVENTION

30 The present invention is directed to po-
tent and selective PDE4 inhibitors useful in treat-
ment of diseases and conditions where inhibition of
PDE4 activity is considered beneficial. The present
PDE4 inhibitors unexpectedly reduce or eliminate the

adverse CNS side effects associated with prior PDE4 inhibitors.

In particular, the present invention is directed to compounds having the structural formula

(II):



(II)

wherein R¹ is lower alkyl, bridged alkyl (e.g., norbornyl), aryl, heteroaryl, aralkyl, cycloalkyl (e.g., indanyl), a 5- or 6-membered saturated heterocycle (e.g., 3-tetrahydrofuryl), C₁₋₄alkylenearyl, C₁₋₄alkyleneOaryl, C₁₋₄alkyleneheteroaryl, C₁₋₄alkyleneHet, C₂₋₄alkylenearyloaryl, C₁₋₄alkylene bridged alkyl, C₁₋₃alkylenecycloalkyl (e.g., cyclopentylmethyl), substituted or unsubstituted propargyl (e.g., -CH₂C≡C-C₆H₅), substituted or unsubstituted allyl (e.g., -CH₂CH=CH-C₆H₅), or halocycloalkyl (e.g., fluorocyclopentyl);

R² is hydrogen, methyl, or halo-substituted methyl, e.g., CHF₂;

R³ is selected from the group consisting of C(=O)OR⁷, NHC(=O)OR⁷, C(=O)R⁷, C(=NH)NR⁸R⁹, C(=O)NR⁸R⁹, lower alkyl, bridged alkyl, cycloalkyl, haloalkyl, halocycloalkyl, C₁₋₃alkylenecycloalkyl, a 5- or 6-

membered saturated heterocycle, aryl, heteroaryl,
C₁₋₃alkyleneC(=O)R⁷, C(=O)C(=O)NR⁸R⁹, C₁₋₄alkyleneOR⁷,
C₁₋₃alkylenearyl, SO₂heteroaryl, Het, aralkyl, alk-
aryl, heteroaralkyl, heteroalkaryl, C₁₋₃alkyleneC-
5 (=O)OR⁷, C(=O)C₁₋₃alkyleneC(=O)OR⁷, C₁₋₃alkylenehetero-
oaryl, C(=O)C(=O)OR⁷, C₁₋₃alkylenearyl, SO₂heteroaryl,
Het, C(=O)C₁₋₃alkyleneC(=O)OR⁷, C(=O)C₁₋₃alkylene-
NH(C=O)OR⁷, C(=O)C₁₋₃alkyleneNH₂, and NHC(=O)OR⁷;

R⁴ is hydrogen, lower alkyl, haloalkyl,
10 cycloalkyl, or aryl;

R⁵ is hydrogen, lower alkyl, alkynyl, halo-
alkyl, cycloalkyl, or aryl;

R⁶ and R¹², independently, are hydrogen,
lower alkyl, aralkyl, SO₂R¹¹, or C(=O)R⁷;

15 R⁷ is selected from the group consisting of
branched or unbranched lower alkyl, heteroaryl, a
heterocycle, aralkyl, and aryl, and R⁷ can be option-
ally substituted with one or more of OR⁸, NR⁸R⁹, or
SR⁸;

20 R⁸ and R⁹, same or different, are selected
from the group consisting of hydrogen, lower alkyl,
cycloalkyl, aryl, heteroaryl, alkaryl, heteroar-
alkyl, heteroalkaryl, and aralkyl, or R⁸ and R⁹ can
be taken together form a 4-membered to 7-membered
25 ring;

R¹⁰ is hydrogen, alkyl, haloalkyl, cyclo-
alkyl, aryl, C(=O)alkyl, C(=O)cycloalkyl, C(=O)aryl,
C(=O)Oalkyl, C(=O)Ocycloalkyl, C(=O)aryl, CH₂OH,
CH₂Oalkyl, CHO, CN, NO₂, or SO₂R¹¹;

30 R¹¹ is alkyl, cycloalkyl, trifluoromethyl,
aryl, aralkyl, or NR⁸R⁹; and

salts and solvates (e.g., hydrates)
thereof.

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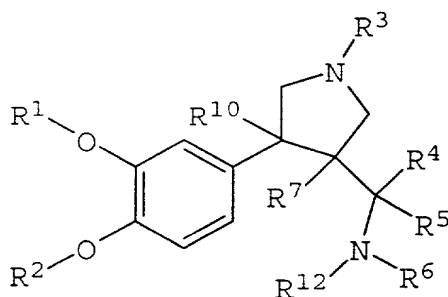
The present invention also is directed to pharmaceutical compositions containing one or more of the compounds of structural formula (II), to use of the compounds and compositions containing the
5 compounds in the treatment of a disease or disorder, and to methods of preparing compounds and intermediates involved in the synthesis of the compounds of structural formula (II).

The present invention also is directed to
10 methods of treating a mammal having a condition where inhibition of PDE4 provides a benefit, modulating cAMP levels in a mammal, reducing TNF α levels in a mammal, suppressing inflammatory cell activation in a mammal, and inhibiting PDE4 function in a
15 mammal by administering therapeutically effective amounts of a compound of structural formula (II), or a composition containing a composition of structural formula (II) to the mammal.

20 DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

The present invention is directed to compounds having the structural formula (II):

5



10

(II)

wherein R¹ is lower alkyl, bridged alkyl (e.g., norbornyl), aryl, heteroaryl, aralkyl, cycloalkyl (e.g., indanyl), a 5- or 6-membered saturated heterocycle (e.g., 3-tetrahydrofuryl), C₁₋₄alkylene-
 15 aryl, C₁₋₄alkyleneOaryl, C₁₋₄alkyleneheteroaryl, C₁₋₄alkyleneHet, C₂₋₄alkylenearyloaryl, C₁₋₄alkylene bridged alkyl, C₁₋₃alkylenecycloalkyl (e.g., cyclopentylmethyl), substituted or unsubstituted
 20 propargyl (i.e., -CH₂C≡C-C₆H₅), substituted or unsubstituted allyl (e.g., -CH₂CH=CH-C₆H₅), or halocycloalkyl (e.g., fluorocyclopentyl);

R² is hydrogen, methyl, or halo-substituted methyl, e.g., CHF₂;

25

R³ is selected from the group consisting of C(=O)OR⁷, NHC(=O)OR⁷, C(=O)R⁷, C(=NH)NR⁸R⁹, C(=O)NR⁸R⁹, lower alkyl, bridged alkyl, cycloalkyl, haloalkyl, halocycloalkyl, C₁₋₃alkylenecycloalkyl, a 5- or 6-membered saturated heterocycle, aryl, heteroaryl,
 30 C₁₋₃alkyleneC(=O)R⁷, C(=O)C(=O)NR⁸R⁹, C₁₋₄alkyleneOR⁷, C₁₋₃alkylenearyl, SO₂heteroaryl, Het, aralkyl, alkaryl, heteroaralkyl, heteroalkaryl, C₁₋₃alkyleneC(=O)OR⁷, C(=O)C₁₋₃alkyleneC(=O)OR⁷, C₁₋₃alkylenehetero-

aryl, $C(=O)C(=O)OR^7$, $C(=O)C_{1-3}alkyleneC(=O)OR^7$, $C(=O)-C_{1-3}alkyleneNH(C=O)OR^7$, $C(=O)C_{1-3}alkyleneNH_2$, and $NHC(=O)OR^7$;

5 R^4 is hydrogen, lower alkyl, haloalkyl, cycloalkyl, or aryl;

R^5 is hydrogen, lower alkyl, alkynyl, haloalkyl, cycloalkyl, or aryl;

R^6 and R^{12} , independently, are hydrogen, lower alkyl, aralkyl, SO_2R^{11} , or $C(=O)R^7$;

10 R^7 is selected from the group consisting of branched or unbranched lower alkyl, heteroaryl, a heterocycle, aralkyl, and aryl, and R^7 can be optionally substituted with one or more of OR^8 , NR^8R^9 , or SR^8 ;

15 R^8 and R^9 , same or different, are selected from the group consisting of hydrogen, lower alkyl, cycloalkyl, aryl, heteroaryl, alkaryl, heteroaralkyl, heteroalkaryl, and aralkyl, or R^8 and R^9 can be taken together form a 4-membered to 7-membered
20 ring;

R^{10} is hydrogen, alkyl, haloalkyl, cycloalkyl, aryl, $C(=O)alkyl$, $C(=O)cycloalkyl$, $C(=O)aryl$, $C(=O)Oalkyl$, $C(=O)Ocycloalkyl$, $C(=O)aryl$, CH_2OH , $CH_2Oalkyl$, CHO , CN , NO_2 , or SO_2R^{11} ;

25 R^{11} is alkyl, cycloalkyl, trifluoromethyl, aryl, aralkyl, or NR^8R^9 ; and

salts and solvates (e.g., hydrates) thereof.

30 As used herein, the term "alkyl," alone or in combination, is defined to include straight chain and branched chain saturated hydrocarbon groups containing one to 16 carbon atoms. The term "lower alkyl" is defined herein as an alkyl group having

one through six carbon atoms (C_1 - C_6). Examples of lower alkyl groups include, but are not limited to, methyl, ethyl, n-propyl, isopropyl, isobutyl, n-butyl, neopentyl, n-hexyl, and the like. The term
5 "alkynyl" refers to an unsaturated alkyl group that contains a carbon-carbon triple bond.

The term "bridged alkyl" is defined herein as a C_6 - C_{16} bicyclic or polycyclic hydrocarbon group, for example, norboryl, adamantyl, bicyclo[2.2.2]-
10 octyl, bicyclo[2.2.1]heptyl, bicyclo[3.2.1]octyl, or decahydronaphthyl.

The term "cycloalkyl" is defined herein to include cyclic C_3 - C_7 hydrocarbon groups, optionally fused to an aromatic group. Examples of cycloalkyl
15 groups include, but are not limited to, cyclopropyl, cyclobutyl, cyclohexyl, cyclopentyl, indanyl, and tetrahydronaphthyl.

The term "alkylene" refers to an alkyl group having a substituent. For example, the term
20 " C_{1-3} alkylenecycloalkyl" refers to an alkyl group containing one to three carbon atoms, and substituted with a cycloalkyl group.

The term "haloalkyl" is defined herein as an alkyl group substituted with one or more halo
25 substituents, either fluoro, chloro, bromo, iodo, or combinations thereof. Similarly, "halocycloalkyl" is defined as a cycloalkyl group having one or more halo substituents.

The term "aryl," alone or in combination,
30 is defined herein as a monocyclic or polycyclic aromatic group, preferably a monocyclic or bicyclic aromatic group, e.g., phenyl or naphthyl, that can be unsubstituted or substituted, for example, with

one or more, and in particular one to three, substituents selected from halo, alkyl, phenyl, hydroxy, hydroxyalkyl, alkoxy, alkoxyalkyl, haloalkyl, nitro, amino, alkylamino, acylamino, alkylthio, alkylsulfinyl, and alkylsulfonyl. Exemplary aryl groups include phenyl, naphthyl, tetrahydronaphthyl, 2-chlorophenyl, 3-chlorophenyl, 4-chlorophenyl, 2-methylphenyl, 4-methoxyphenyl, 3-trifluoromethyl-phenyl, 4-nitrophenyl, and the like.

10 The term "heteroaryl" is defined herein as a monocyclic or bicyclic ring system containing one or two aromatic rings and containing at least one nitrogen, oxygen, or sulfur atom in an aromatic ring, and which can be unsubstituted or substituted, for example, with one or more, and in particular one to three, substituents, like halo, alkyl, hydroxy, hydroxyalkyl, alkoxy, alkoxyalkyl, haloalkyl, nitro, amino, alkylamino, acylamino, alkylthio, alkylsulfinyl, and alkylsulfonyl. Examples of heteroaryl groups include thienyl, furyl, pyridyl, oxazolyl, quinolyl, isoquinolyl, indolyl, triazolyl, isothiazolyl, isoxazolyl, imidazolyl, benzothiazolyl, pyrazinyl, pyrimidinyl, thiazolyl, and thiadiazolyl.

20 The term "aralkyl" is defined herein as a previously defined alkyl group, wherein one of the hydrogen atoms is replaced by an aryl group as defined herein, for example, a phenyl group optionally having one or more substituents, for example, halo, alkyl, alkoxy, and the like. An example of an aralkyl group is a benzyl group.

25 The term "alkaryl" is defined herein as a previously defined aryl group, wherein one of the

hydrogen atoms is replaced by an alkyl, cycloalkyl, haloalkyl, or halocycloalkyl group.

The terms "heteroaralkyl" and "hetero-alkaryl" are defined similarly as the term "aralkyl" and "alkaryl," however, the aryl group is replaced by a heteroaryl group as previously defined.

The term "heterocycle" is defined as a 5- or 6-membered nonaromatic ring having one or more heteroatoms selected from oxygen, nitrogen, and sulfur present in the ring. Nonlimiting examples include tetrahydrofuran, piperidine, piperazine, sulfolane, morpholine, tetrahydropyran, dioxane, and the like.

The term "halogen" or "halo" is defined herein to include fluorine, chlorine, bromine, and iodine.

The terms "alkoxy," "aryloxy," and "aralkoxy" are defined as -OR, wherein R is alkyl, aryl, and aralkyl, respectively.

The term "alkoxyalkyl" is defined as an alkoxy group appended to an alkyl group. The terms "aryloxyalkyl" and "aralkoxyalkyl" are similarly defined as an aryloxy or aralkoxy group appended to an alkyl group.

The term "hydroxy" is defined as -OH.

The term "hydroxyalkyl" is defined as a hydroxy group appended to an alkyl group.

The term "amino" is defined as -NH₂.

The term "alkylamino" is defined as -NR₂ wherein at least one R is alkyl and the second R is alkyl or hydrogen.

The term "acylamino" is defined as RC(=O)N, wherein R is alkyl or aryl.

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The term "nitro" is defined as $-\text{NO}_2$.

The term "alkylthio" is defined as $-\text{SR}$,
where R is alkyl.

The term "alkylsulfinyl" is defined as
5 $\text{R}-\text{SO}_2$, where R is alkyl.

The term "alkylsulfonyl" is defined as
 $\text{R}-\text{SO}_3$, where R is alkyl.

In preferred embodiments, R^5 is hydrogen or
methyl, R^7 is methyl, R^2 is methyl or difluoromethyl,
10 R^4 is selected from the group consisting of hydrogen,
lower alkyl, benzyl, and phenyl, R^{12} is hydrogen or
methyl, and R^6 is selected from the group consisting
of hydrogen, methyl, ethyl, $\text{C}(=\text{O})\text{R}^7$, $\text{C}(=\text{O})\text{OR}^7$,
benzyl, SO_2CH_3 , and $\text{SO}_2\text{C}_6\text{H}_5$. R^1 is selected from the
15 group consisting of

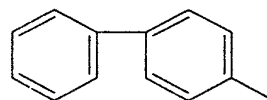
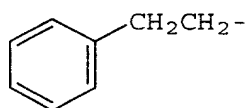
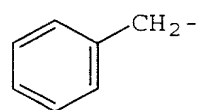
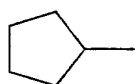
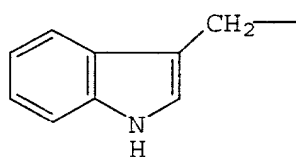
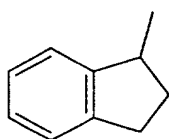
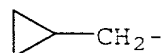
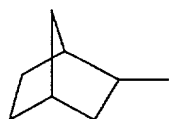
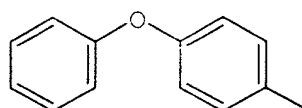
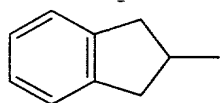
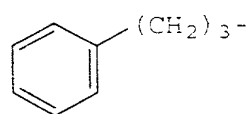
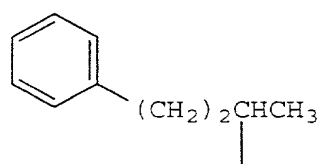
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Figure 6

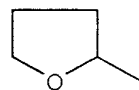
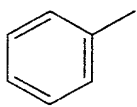
Figure 6 displays six histograms showing the distribution of the number of nodes per cluster for different values of α . The x-axis represents the number of nodes per cluster, ranging from 0 to 10. The y-axis represents the frequency or probability density. The distributions are labeled as follows:

- (a) $\alpha = 0.0$: Shows a highly skewed distribution with a peak at 1 node.
- (b) $\alpha = 0.1$: Shows a more spread-out distribution compared to (a).
- (c) $\alpha = 0.2$: Shows a distribution shifted towards higher numbers of nodes per cluster.
- (d) $\alpha = 0.3$: Shows a distribution shifted further towards higher numbers of nodes per cluster.
- (e) $\alpha = 0.4$: Shows a distribution shifted even further towards higher numbers of nodes per cluster.
- (f) $\alpha = 0.5$: Shows a distribution shifted towards higher numbers of nodes per cluster, with a peak around 4-5 nodes.

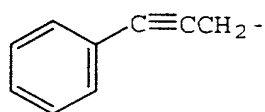
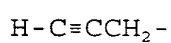
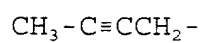
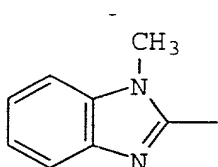
The figure illustrates how increasing α affects the clustering behavior, leading to larger clusters.



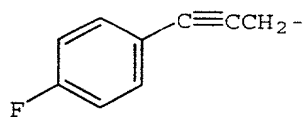
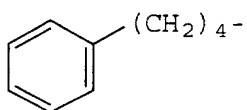
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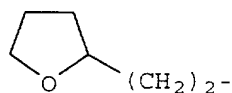
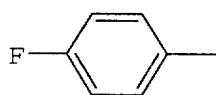
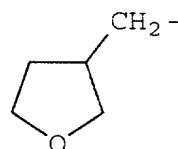
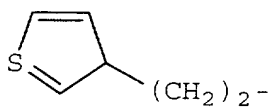
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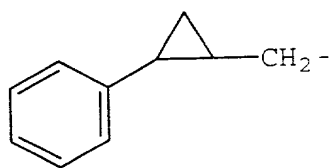


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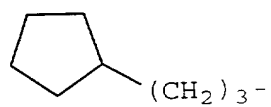


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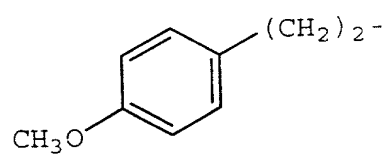
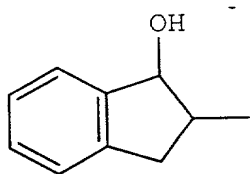
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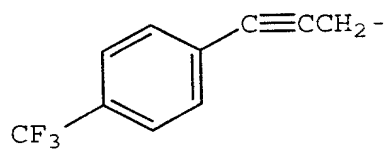
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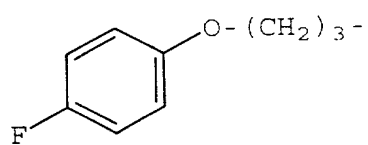
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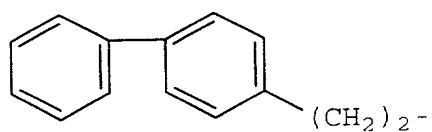
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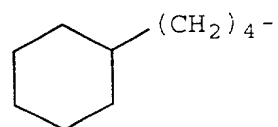
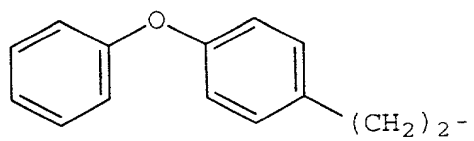
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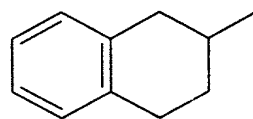
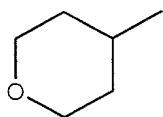


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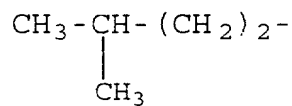
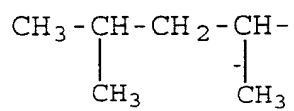


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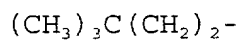
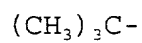




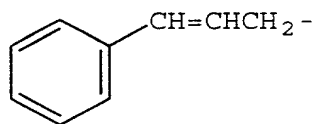
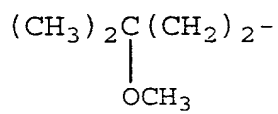
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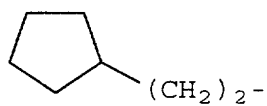
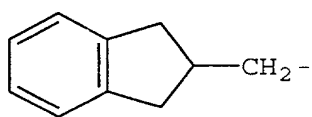
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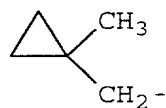
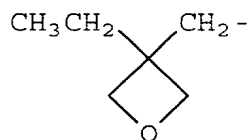


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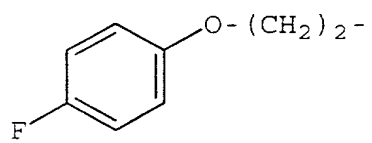


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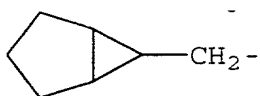
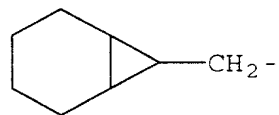
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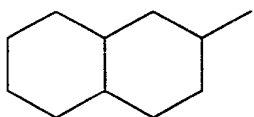
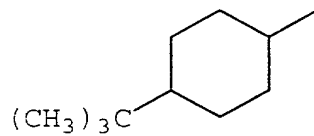
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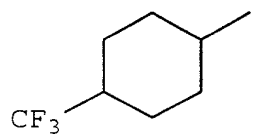
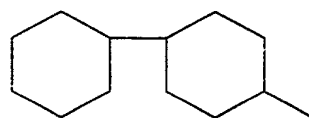
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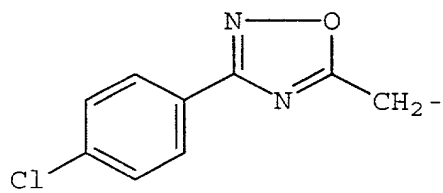
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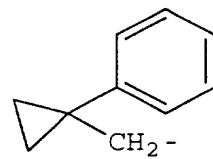
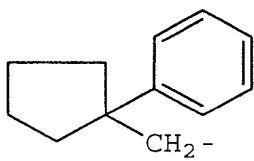
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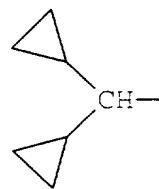
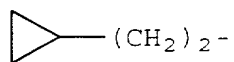


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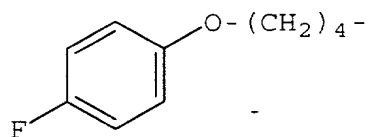


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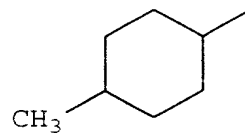
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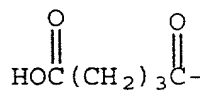
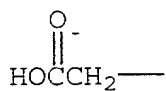
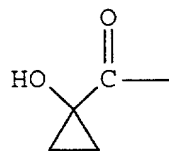
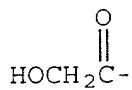
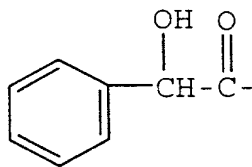
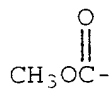
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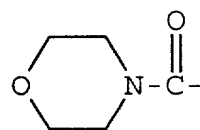
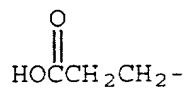
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R³ is selected from the group consisting of

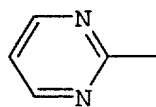
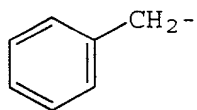
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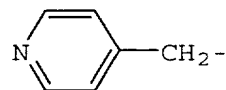
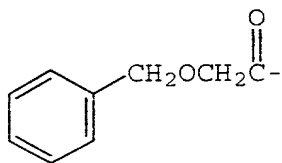
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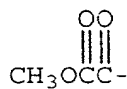
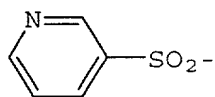
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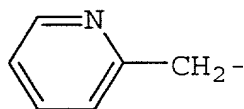
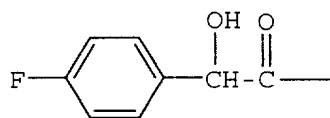
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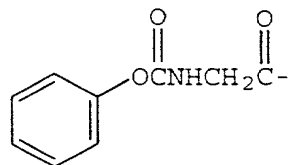
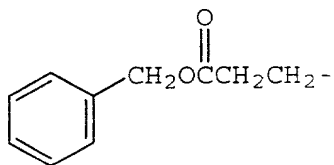


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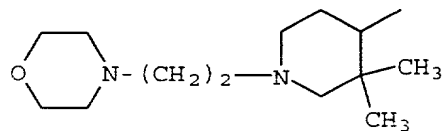
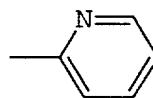
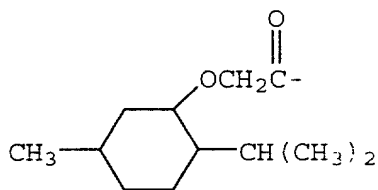
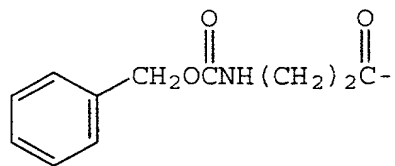
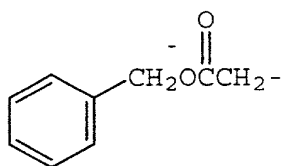


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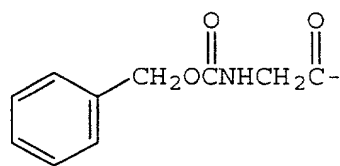
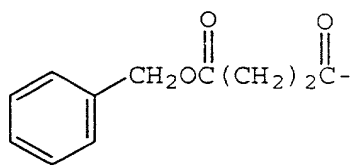
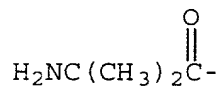
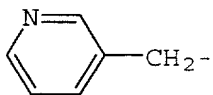




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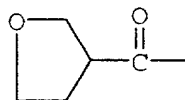
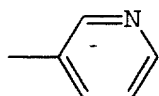
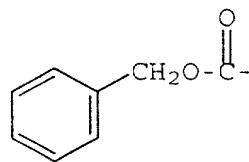
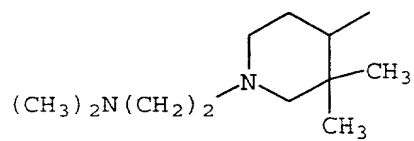


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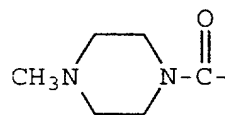
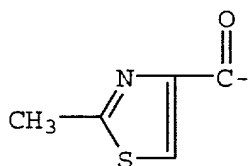


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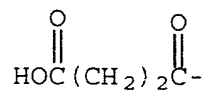
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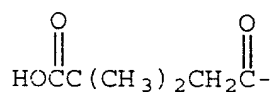
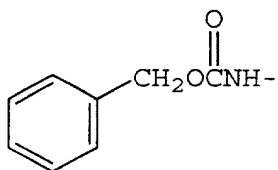
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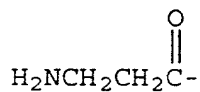
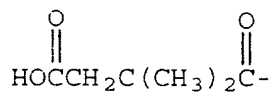
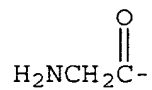
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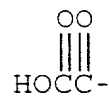
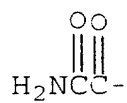


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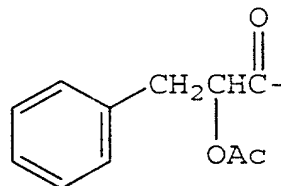
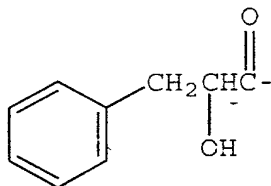


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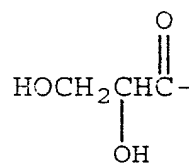
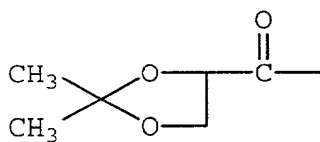
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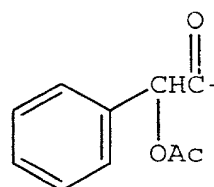
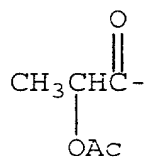
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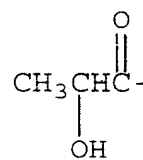
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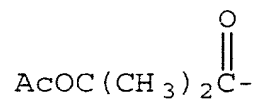
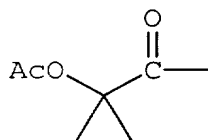
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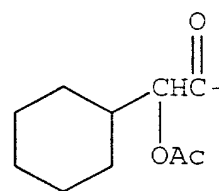
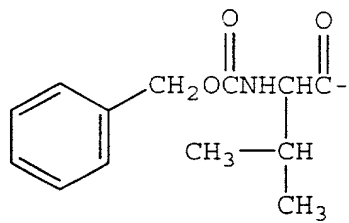
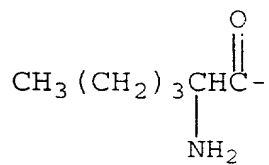
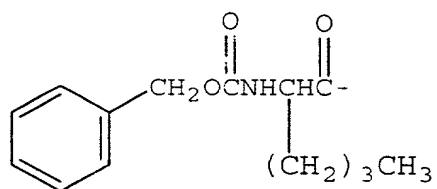
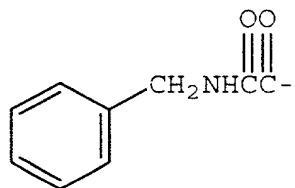
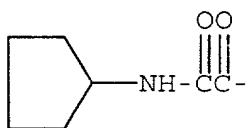
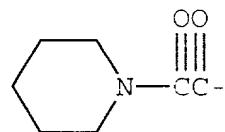
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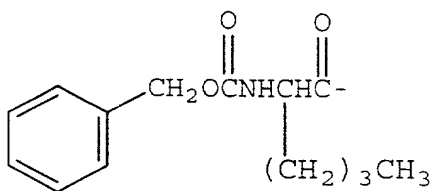
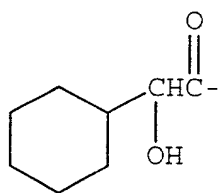
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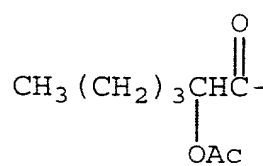
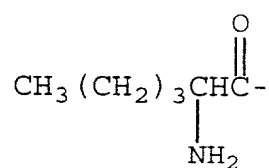
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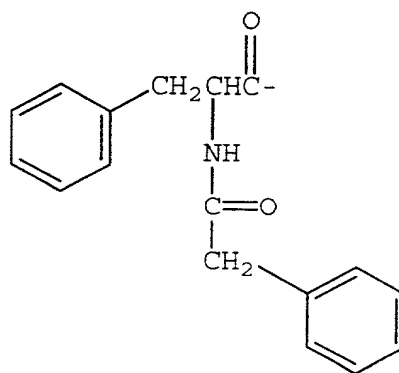
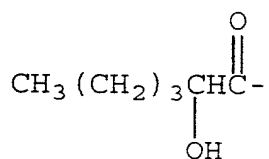
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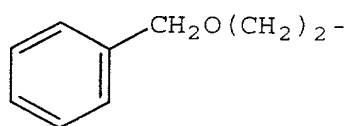
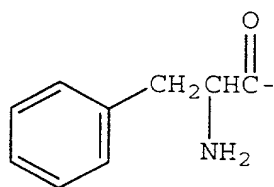


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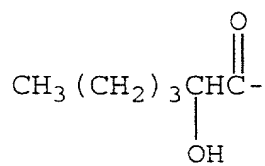
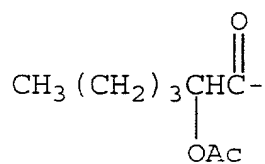
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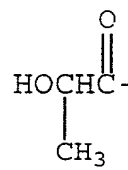
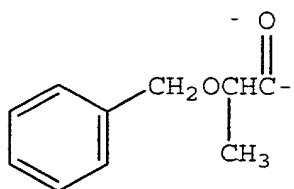


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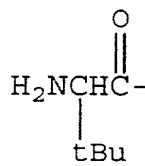
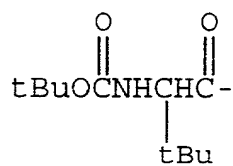
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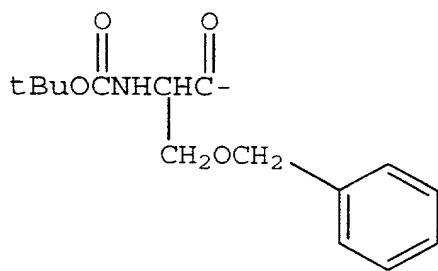
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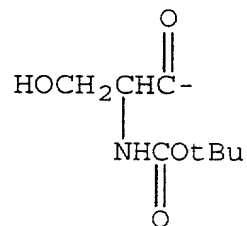


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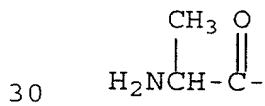
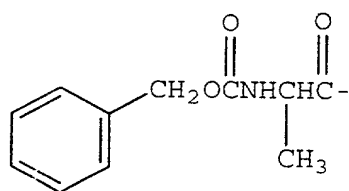
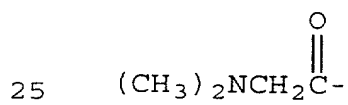
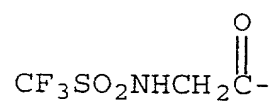
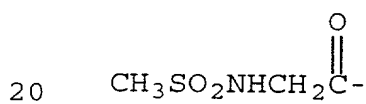
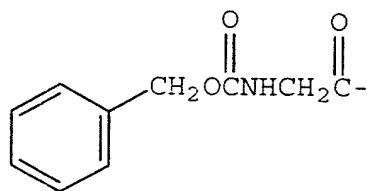
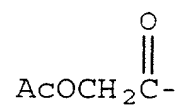
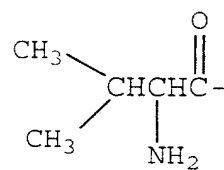
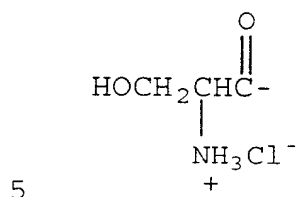


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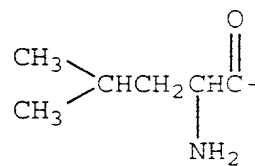
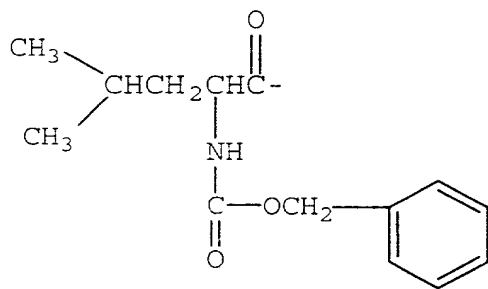
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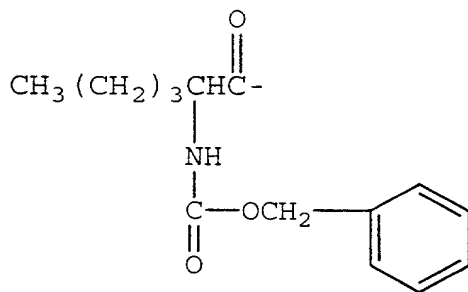
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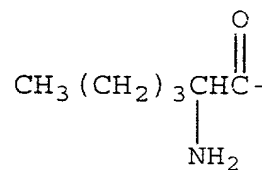
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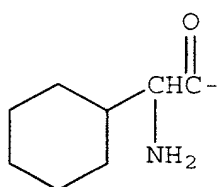
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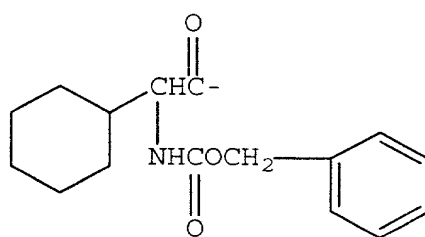
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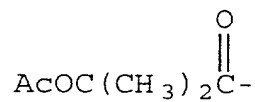
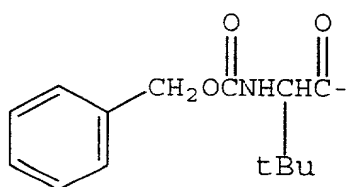
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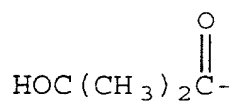
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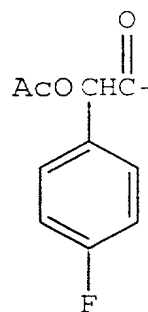
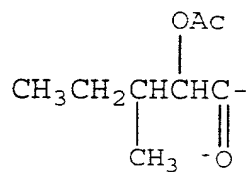
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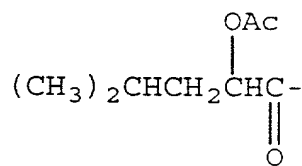
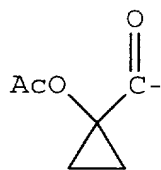
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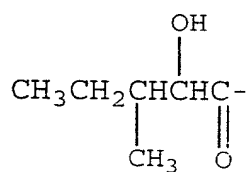
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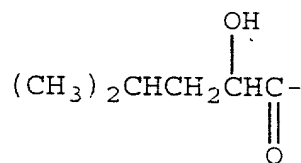
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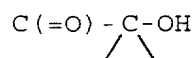
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wherein Ac is $\text{CH}_3\text{C}(=\text{O})$ and tBu is $\text{C}(\text{CH}_3)_3$.

25

In most preferred embodiments, R¹ is selected from the group consisting of cyclopentyl, cyclopropylmethyl, tetrahydrofuryl, indanyl, norbornyl, phenethyl, and phenylbutyl; R² is selected from the group consisting of methyl and difluoromethyl; R³ is selected from the group consisting of benzyl, CO₂CH₃, C(=O)CH₂OH, C(=O)CH(CH₃)OH, C(=O)C(CH₃)₂OH, and

10



;

15 R⁴ is hydrogen; R⁵ is hydrogen or methyl; R⁶ is hydrogen, methyl, ethyl, benzyl, SO₂CH₃, SO₂C₆H₅, benzoyl, C(=O)C(CH₃)₃, or acetyl; R¹² is hydrogen or methyl; and R¹⁰ is hydrogen.

The present invention includes all possible stereoisomers and geometric isomers of compounds of structural formula (II), and includes not only racemic compounds but also the optically active isomers as well. When a compound of structural formula (II) is desired as a single enantiomer, it can be obtained either by resolution of the final product or by stereospecific synthesis from either isomerically pure starting material or use of a chiral auxiliary reagent, for example, see Z. Ma et al., *Tetrahedron: Asymmetry*, 8(6), pages 883-888 (1997). Resolution of the final product, an intermediate, or a starting material can be achieved by any suitable method known in the art. Additionally, in situations where tautomers of the compounds of

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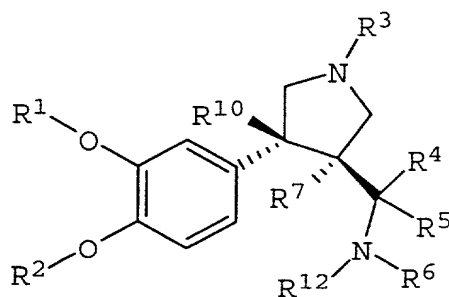
structural formula (II) are possible, the present invention is intended to include all tautomeric forms of the compounds. As demonstrated hereafter, specific stereoisomers exhibit an exceptional ability to inhibit PDE4 without manifesting the adverse CNS side effects typically associated with PDE4 inhibitors.

In particular, it is generally accepted that biological systems can exhibit very sensitive activities with respect to the absolute stereochemical nature of compounds. (See, E.J. Ariens, *Medicinal Research Reviews*, 6:451-466 (1986); E.J. Ariens, *Medicinal Research Reviews*, 7:367-387 (1987); K.W. Fowler, *Handbook of Stereoisomers: Therapeutic Drugs*, CRC Press, edited by Donald P. Smith, pp. 35-63 (1989); and S.C. Stinson, *Chemical and Engineering News*, 75:38-70 (1997).)

For example, rolipram is a stereospecific PDE4 inhibitor that contains one chiral center. The (-)-enantiomer of rolipram has a higher pharmacological potency than the (+)-enantiomer, which could be related to its potential antidepressant action. Schultz et al., *Naunyn-Schmiedeberg's Arch Pharmacol*, 333:23-30 (1986). Furthermore, the metabolism of rolipram appears stereospecific with the (+)-enantiomer exhibiting a faster clearance rate than the (-)-enantiomer. Krause et al., *Xenobiotica*, 18:561-571 (1988). Finally, a recent observation indicated that the (-)-enantiomer of rolipram (R-rolipram) is about ten-fold more emetic than the (+)-enantiomer (S-rolipram). A. Robichaud et al., *Neuropharmacology*, 38:289-297 (1999). This observation is not easily reconciled with differences in

test animal disposition to rolipram isomers and the ability of rolipram to inhibit the PDE4 enzyme. The compounds of the present invention can have three or more chiral centers. As shown below, compounds of a
5 specific stereochemical orientation exhibit similar PDE4 inhibitory activity and pharmacological activity, but altered CNS toxicity and emetic potential.

Accordingly, preferred compounds of the present invention have the structural formula (III):



(III)

The compounds of structural formula (III) are potent and selective PDE4 inhibitors, and do not manifest the adverse CNS effects and emetic potential demonstrated by stereoisomers of a compound of structural
25 formula (III).

Compounds of structural formula (II) which contain acidic moieties can form pharmaceutically acceptable salts with suitable cations. Suitable
30 pharmaceutically acceptable cations include alkali metal (e.g., sodium or potassium) and alkaline earth metal (e.g., calcium or magnesium) cations. The pharmaceutically acceptable salts of the compounds

of structural formula (II), which contain a basic center, are acid addition salts formed with pharmaceutically acceptable acids. Examples include the hydrochloride, hydrobromide, sulfate or bisulfate, phosphate or hydrogen phosphate, acetate, benzoate, succinate, fumarate, maleate, lactate, citrate, tartrate, gluconate, methanesulfonate, benzene-sulphonate, and p-toluenesulphonate salts. In light of the foregoing, any reference to compounds of the present invention appearing herein is intended to include compounds of structural formula (II), as well as pharmaceutically acceptable salts and solvates thereof.

The compounds of the present invention can be therapeutically administered as the neat chemical, but it is preferable to administer compounds of structural formula (II) as a pharmaceutical composition or formulation. Accordingly, the present invention further provides for pharmaceutical formulations comprising a compound of structural formula (II), together with one or more pharmaceutically acceptable carriers and, optionally, other therapeutic and/or prophylactic ingredients. The carriers are "acceptable" in the sense of being compatible with the other ingredients of the formulation and not deleterious to the recipient thereof.

In particular, a selective PDE4 inhibitor of the present invention is useful alone or in combination with a second antiinflammatory therapeutic agent, for example, a therapeutic agent targeting TNF α , such as ENBREL[®] or REMICADE[®], which have utility in treating rheumatoid arthritis. Likewise, therapeutic utility of IL-1 antagonism has also been

shown in animal models for rheumatoid arthritis. Thus, it is envisioned that IL-1 antagonism, in combination with PDE4 inhibition, which attenuates TNF α , would be efficacious.

5 The present PDE4 inhibitors are useful in the treatment of a variety of allergic, autoimmune, and inflammatory diseases.

 The term "treatment" includes preventing, lowering, stopping, or reversing the progression of
10 severity of the condition or symptoms being treated. As such, the term "treatment" includes both medical therapeutic and/or prophylactic administration, as appropriate.

 In particular, inflammation is a local-
15 ized, protective response elicited by injury or destruction of tissues, which serves to destroy, dilute or wall off (i.e., sequester) both the injurious agent and the injured tissue. The term "inflammatory disease," as used herein, means any dis-
20 ease in which an excessive or unregulated inflammatory response leads to excessive inflammatory symptoms, host tissue damage, or loss of tissue function. Additionally, the term "autoimmune disease," as used herein, means any group of disorders in
25 which tissue injury is associated with humoral or cell-mediated responses to the body's own constituents. The term "allergic disease," as used herein, means any symptoms, tissue damage, or loss of tissue function resulting from allergy. The term "arth-
30 ritic disease," as used herein, means any of a large family of diseases that are characterized by inflammatory lesions of the joints attributable to a variety of etiologies. The term "dermatitis," as used

herein, means any of a large family of diseases of the skin that are characterized by inflammation of the skin attributable to a variety of etiologies. The term "transplant rejection," as used herein, means any immune reaction directed against grafted tissue (including organ and cell (e.g., bone marrow)), characterized by a loss of function of the grafted and surrounding tissues, pain, swelling, leukocytosis and thrombocytopenia.

10 The present invention also provides a method of modulating cAMP levels in a mammal, as well as a method of treating diseases characterized by elevated cytokine levels.

15 The term "cytokine," as used herein, means any secreted polypeptide that affects the functions of other cells, and that modulates interactions between cells in the immune or inflammatory response. Cytokines include, but are not limited to monokines, lymphokines, and chemokines regardless of which cells produce them. For instance, a monokine is generally referred to as being produced and secreted by a monocyte, however, many other cells produce monokines, such as natural killer cells, fibroblasts, basophils, neutrophils, endothelial cells, brain astrocytes, bone marrow stromal cells, epidermal keratinocytes, and B-lymphocytes. Lymphokines are generally referred to as being produced by lymphocyte cells. Examples of cytokines include, but are not limited to, interleukin-1 (IL-1), interleukin-6 (IL-6), Tumor Necrosis Factor alpha (TNF α), and Tumor Necrosis Factor beta (TNF β).

30 The present invention further provides a method of reducing TNF levels in a mammal, which

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comprises administering an effective amount of a compound of structural formula (II) to the mammal. The term "reducing TNF levels," as used herein, means either:

- 5 a) decreasing excessive *in vivo* TNF levels in a mammal to normal levels or below normal levels by inhibition of the *in vivo* release of TNF by all cells, including but not limited to monocytes or macrophages; or
- 10 b) inducing a down-regulation, at the translational or transcription level, of excessive *in vivo* TNF levels in a mammal to normal levels or below normal levels; or
- 15 c) inducing a down-regulation, by inhibition of the direct synthesis of TNF as a postranslational event.

Moreover, the compounds of the present invention are useful in suppressing inflammatory cell activation. The term "inflammatory cell activation," as used herein, means the induction by a stimulus (including, but not limited to, cytokines, antigens or auto-antibodies) of a proliferative cellular response, the production of soluble mediators (including but not limited to cytokines, oxygen radicals, enzymes, prostanoids, or vasoactive amines), or cell surface expression of new or increased numbers of mediators (including, but not limited to, major histocompatibility antigens or cell adhesion molecules) in inflammatory cells (including but not limited to monocytes, macrophages, T lymphocytes, B lymphocytes, granulocytes, polymorphonuclear leukocytes, mast cells, basophils, eosinophils, dendritic cells, and endothelial

cells). It will be appreciated by persons skilled in the art that the activation of one or a combination of these phenotypes in these cells can contribute to the initiation, perpetuation, or exacerbation of an inflammatory condition.

The compounds of the present invention also are useful in causing airway smooth muscle relaxation, bronchodilation, and prevention of bronchoconstriction.

10 The compounds of the present invention, therefore, are useful in treating such diseases as arthritic diseases (such as rheumatoid arthritis), osteoarthritis, gouty arthritis, spondylitis, thyroid-associated ophthalmopathy, Behcet disease, 15 sepsis, septic shock, endotoxic shock, gram negative sepsis, gram positive sepsis, toxic shock syndrome, asthma, chronic bronchitis, allergic rhinitis, allergic conjunctivitis, vernal conjunctivitis, eosinophilic granuloma, adult (acute) respiratory 20 distress syndrome (ARDS), chronic pulmonary inflammatory disease (such as chronic obstructive pulmonary disease), silicosis, pulmonary sarcoidosis, reperfusion injury of the myocardium, brain or extremities, brain or spinal cord injury due to minor 25 trauma, fibrosis including cystic fibrosis, keloid formation, scar tissue formation, atherosclerosis, autoimmune diseases, such as systemic lupus erythematosus (SLE) and transplant rejection disorders (e.g., graft vs. host (GvH) reaction and allo- 30 graft rejection), chronic glomerulonephritis, inflammatory bowel diseases, such as Crohn's disease and ulcerative colitis, proliferative lymphocytic diseases, such as leukemias (e.g. chronic lympho-

cytic leukemia; CLL) (see Mentz et al., *Blood* 88, pp. 2172-2182 (1996)), and inflammatory dermatoses, such as atopic dermatitis, psoriasis, or urticaria.

Other examples of such diseases or related conditions include cardiomyopathies, such as congestive heart failure, pyrexia, cachexia, cachexia secondary to infection or malignancy, cachexia secondary to acquired immune deficiency syndrome (AIDS), ARC (AIDS-related complex), cerebral malaria, osteoporosis and bone resorption diseases, and fever and myalgias due to infection. In addition, the compounds of the present invention are useful in the treatment of diabetes insipidus and central nervous system disorders, such as depression and multi-infarct dementia.

Compounds of the present invention also have utility outside of that typically known as therapeutic. For example, the present compounds can function as organ transplant preservatives (see Pinsky et al., *J. Clin. Invest.*, 92, pp. 2994-3002 (1993)) as well.

Selective PDE4 inhibitors also can be useful in the treatment of erectile dysfunction, especially vasculogenic impotence (Doherty, Jr. et al. U.S. Patent No. 6,127,363), diabetes insipidus (*Kidney Int.*, 37, p. 362 (1990); *Kidney Int.*, 35, p. 494, (1989)) and central nervous system disorders, such as multiinfarct dementia (Nicholson, *Psychopharmacology*, 101, p. 147 (1990)), depression (Eckman et al., *Curr. Ther. Res.*, 43, p. 291 (1988)), anxiety and stress responses (*Neuropharmacology*, 38, p. 1831 (1991)), cerebral ischemia (*Eur. J. Pharmacol.*, 272, p. 107 (1995)), tardive dys-

kinesia (*J. Clin. Pharmacol.*, 16, p. 304 (1976)),
Parkinson's disease (see *Neurology*, 25, p. 722
(1975); *Clin. Exp. Pharmacol, Physiol.*, 26, p. 421
(1999)), and premenstrual syndrome. With respect to
5 depression, PDE4-selective inhibitors show efficacy
in a variety of animal models of depression such as
the "behavioral despair" or Porsolt tests (*Eur. J.*
Pharmacol., 47, p. 379 (1978); *Eur. J. Pharmacol.*,
57, p. 431 (1979); *Antidepressants: neurochemical,*
10 *behavioral and clinical perspectives*, Enna, Malick,
and Richelson, eds., Raven Press, p. 121 (1981)),
and the "tail suspension test" (*Psychopharmacology*,
85, p. 367 (1985)). Recent research findings show
that chronic *in vivo* treatment by a variety of anti-
15 depressants increase the brain-derived expression of
PDE4 (*J. Neuroscience*, 19, p. 610 (1999)). There-
fore, a selective PDE4 inhibitor can be used alone
or in conjunction with a second therapeutic agent in
a treatment for the four major classes of antide-
20 pressants: electroconvulsive procedures, monoamine
oxidase inhibitors, and selective reuptake inhibi-
tors of serotonin or norepinephrine. Selective PDE4
inhibitors also can be useful in applications that
modulate bronchodilatory activity via direct action
25 on bronchial smooth muscle cells for the treatment
of asthma.

Compounds and pharmaceutical compositions
suitable for use in the present invention include
those wherein the active ingredient is administered
30 to a mammal in an effective amount to achieve its
intended purpose. More specifically, a "therapeuti-
cally effective amount" means an amount effective to
prevent development of, or to alleviate the existing

symptoms of, the subject being treated. Determination of the effective amounts is well within the capability of those skilled in the art, especially in light of the detailed disclosure provided herein.

5 The term "mammal" as used herein includes males and females, and encompasses humans, domestic animals (e.g., cats, dogs), livestock (e.g., cattle, horses, swine), and wildlife (e.g., primates, large cats, zoo specimens).

10 A "therapeutically effective dose" refers to that amount of the compound that results in achieving the desired effect. Toxicity and therapeutic efficacy of such compounds can be determined by standard pharmaceutical procedures in cell cultures or experimental animals, e.g., for determining
15 the LD₅₀ (the dose lethal to 50% of the population) and the ED₅₀ (the dose therapeutically effective in 50% of the population). The dose ratio between toxic and therapeutic effects is the therapeutic
20 index, which is expressed as the ratio between LD₅₀ and ED₅₀. Compounds which exhibit high therapeutic indices are preferred. The data obtained from such data can be used in formulating a dosage range for use in humans. The dosage of such compounds preferably
25 lies within a range of circulating concentrations that include the ED₅₀ with little or no toxicity. The dosage can vary within this range depending upon the dosage form employed, and the route of administration utilized.

30 The exact formulation, route of administration, and dosage can be chosen by the individual physician in view of the patient's condition. Dosage amount and interval can be adjusted individually

to provide plasma levels of the active moiety which are sufficient to maintain the therapeutic effects.

As appreciated by persons skilled in the art, reference herein to treatment extends to prophylaxis, as well as to treatment of established diseases or symptoms. It is further appreciated that the amount of a compound of the invention required for use in treatment varies with the nature of the condition being treated, and with the age and the condition of the patient, and is ultimately determined by the attendant physician or veterinarian. In general, however, doses employed for adult human treatment typically are in the range of 0.001 mg/kg to about 100 mg/kg per day. The desired dose can be conveniently administered in a single dose, or as multiple doses administered at appropriate intervals, for example as two, three, four or more subdoses per day. In practice, the physician determines the actual dosing regimen which is most suitable for an individual patient, and the dosage varies with the age, weight, and response of the particular patient. The above dosages are exemplary of the average case, but there can be individual instances in which higher or lower dosages are merited, and such are within the scope of the present invention.

Formulations of the present invention can be administered in a standard manner for the treatment of the indicated diseases, such as orally, parenterally, transmucosally (e.g., sublingually or via buccal administration), topically, transdermally, rectally, via inhalation (e.g., nasal or deep lung inhalation). Parenteral administration

includes, but is not limited to intravenous, intra-
arterial, intraperitoneal, subcutaneous, intramuscu-
lar, intrathecal, and intraarticular. Parenteral
administration also can be accomplished using a high
5 pressure technique, like POWDERJECT™.

For buccal administration, the composition
can be in the form of tablets or lozenges formulated
in conventional manner. For example, tablets and
capsules for oral administration can contain conven-
10 tional excipients such as binding agents (for exam-
ple, syrup, acacia, gelatin, sorbitol, tragacanth,
mucilage of starch or polyvinylpyrrolidone), fillers
(for example, lactose, sugar, microcrystalline,
cellulose, maize-starch, calcium phosphate or sorbi-
15 tol), lubricants (for example, magnesium, stearate,
stearic acid, talc, polyethylene glycol or silica),
disintegrants (for example, potato starch or sodium
starch glycollate), or wetting agents (for example,
sodium lauryl sulfate). The tablets can be coated
20 according to methods well known in the art.

Alternatively, the compounds of the pres-
ent invention can be incorporated into oral liquid
preparations such as aqueous or oily suspensions,
solutions, emulsions, syrups, or elixirs, for exam-
25 ple. Moreover, formulations containing these com-
pounds can be presented as a dry product for consti-
tution with water or other suitable vehicle before
use. Such liquid preparations can contain conven-
tional additives, such as suspending agents, such as
30 sorbitol syrup, methyl cellulose, glucose/sugar
syrup, gelatin, hydroxyethylcellulose, hydroxy-
propylmethylcellulose, carboxymethyl cellulose,
aluminum stearate gel, and hydrogenated edible fats;

emulsifying agents, such as lecithin, sorbitan mono-
oleate, or acacia; nonaqueous vehicles (which can
include edible oils), such as almond oil, fraction-
ated coconut oil, oily esters, propylene glycol, and
5 ethyl alcohol; and preservatives, such as methyl or
propyl p-hydroxybenzoate and sorbic acid.

Such preparations also can be formulated
as suppositories, e.g., containing conventional
suppository bases, such as cocoa butter or other
10 glycerides. Compositions for inhalation typically
can be provided in the form of a solution, suspen-
sion, or emulsion that can be administered as a dry
powder or in the form of an aerosol using a conven-
tional propellant, such as dichlorodifluoromethane
15 or trichlorofluoromethane. Typical topical and
transdermal formulations comprise conventional aque-
ous or nonaqueous vehicles, such as eye drops,
creams, ointments, lotions, and pastes, or are in
the form of a medicated plaster, patch, or membrane.

20 Additionally, compositions of the present
invention can be formulated for parenteral adminis-
tration by injection or continuous infusion. Formu-
lations for injection can be in the form of suspen-
sions, solutions, or emulsions in oily or aqueous
25 vehicles, and can contain formulation agents, such
as suspending, stabilizing, and/or dispersing
agents. Alternatively, the active ingredient can be
in powder form for constitution with a suitable
vehicle (e.g., sterile, pyrogen-free water) before
30 use.

A composition in accordance with the
present invention also can be formulated as a depot
preparation. Such long acting formulations can be

administered by implantation (for example, subcutaneously or intramuscularly) or by intramuscular injection. Accordingly, the compounds of the invention can be formulated with suitable polymeric or hydrophobic materials (e.g., an emulsion in an acceptable oil), ion exchange resins, or as sparingly soluble derivatives (e.g., a sparingly soluble salt).

For veterinary use, a compound of formula (II), or nontoxic salts thereof, is administered as a suitably acceptable formulation in accordance with normal veterinary practice. The veterinarian can readily determine the dosing regimen and route of administration that is most appropriate for a particular animal.

Thus, the invention provides in a further aspect a pharmaceutical composition comprising a compound of the formula (II), together with a pharmaceutically acceptable diluent or carrier therefor. There is further provided by the present invention a process of preparing a pharmaceutical composition comprising a compound of formula (II), which process comprises mixing a compound of formula (II), together with a pharmaceutically acceptable diluent or carrier therefor.

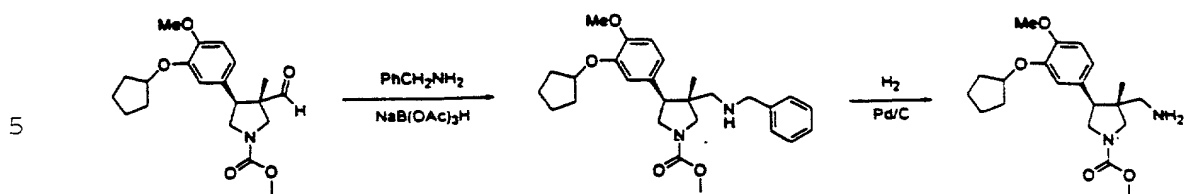
Specific, nonlimiting examples of compounds of structural formula (II) are provided below, the synthesis of which were performed in accordance with the procedures set forth below.

Generally, compounds of structural formula (II) can be prepared according to the following synthetic schemes. In each scheme described below, it is understood in the art that protecting groups

can be employed where necessary in accordance with general principles of synthetic chemistry. These protecting groups are removed in the final steps of the synthesis under basic, acidic, or hydrogenolytic conditions which are readily apparent to those skilled in the art. By employing appropriate manipulation and protection of any chemical functionalities, synthesis of compounds of structural formula (II) not specifically set forth herein can be accomplished by methods analogous to the schemes set forth below.

Unless otherwise noted, all starting materials were obtained from commercial suppliers and used without further purification. All reactions and chromatography fractions were analyzed by thin-layer chromatography on 250-mm silica gel plates, visualized with UV (ultraviolet) or light I_2 (iodine) stain. Products and intermediates were purified by flash chromatography, or reverse-phase HPLC.

The compounds of general structural formula (II) can be prepared, for example, as set forth in the following synthetic scheme. Other synthetic routes also are known to persons skilled in the art. The following reaction scheme provides a compound of structural formula (II), wherein R^1 and R^2 , i.e., CH_3 and cyclopentyl, are determined by the starting materials. Proper selection of other starting materials, or performing conversion reactions on intermediates and examples, provide compounds of general structural formula (II) having other recited R^1 through R^{11} substituents.

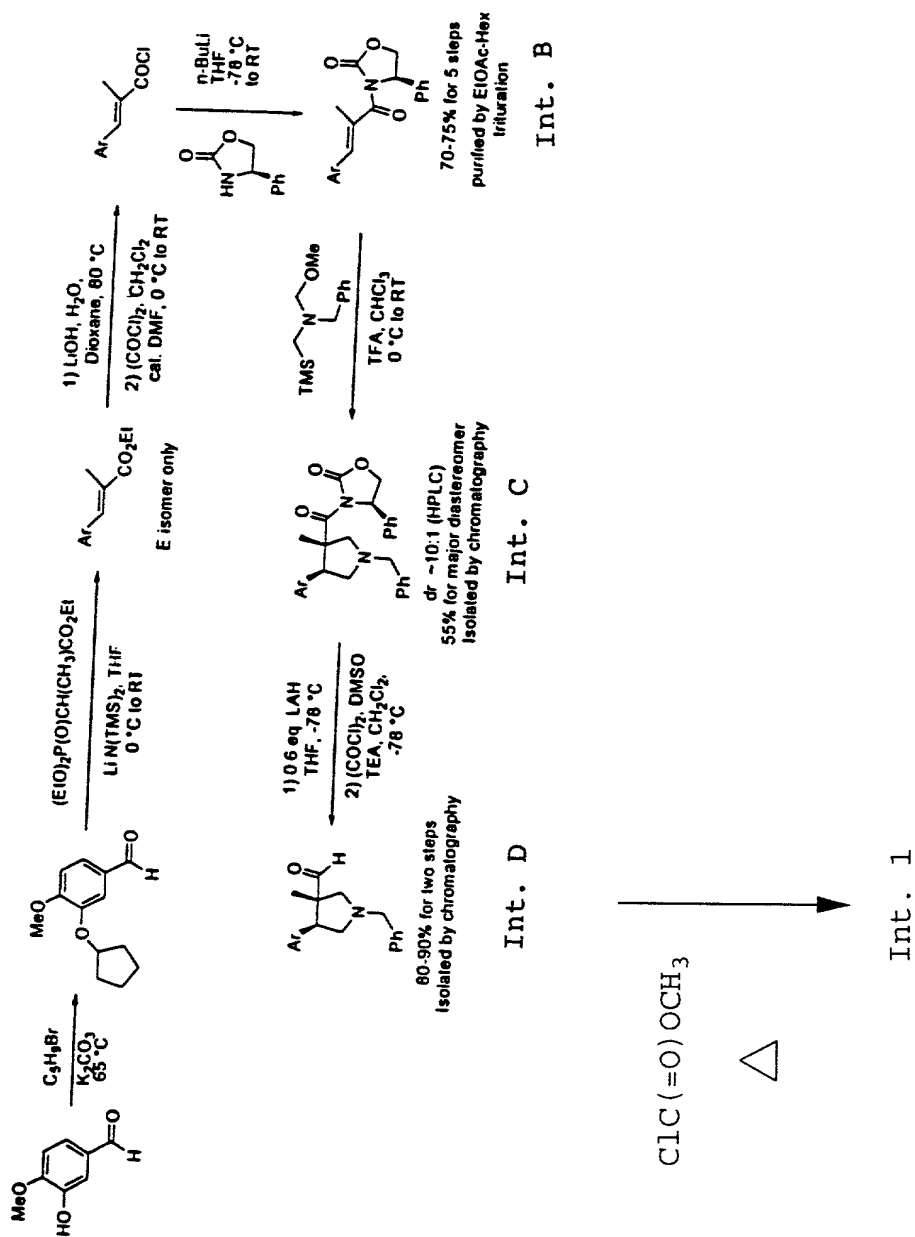


Intermediate 1

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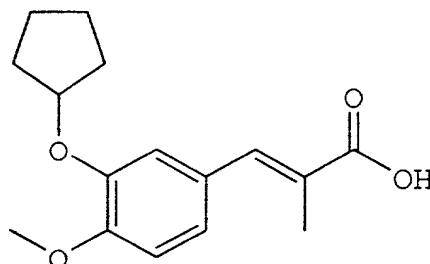
Intermediate 1 was prepared by the following synthetic sequence.



The following illustrates the synthesis of various intermediates and compounds of structural formula (II). The following examples are provided for illustration and should not be construed as limiting.

Intermediate A

Preparation of (E)-3-(3-Cyclopentoxo-4-methoxy-phenyl)-2-methyl-acrylic acid



First, (E)-3-(3-cyclopentoxo-4-methoxyphenyl)-2-methyl-acrylic acid ethyl ester was prepared as follows:

To a cooled (0°C), stirred solution of triethylphosphonopropionate (50.6 mL; 236 mmol; 1.05 eq.) in dry tetrahydrofuran (500 mL) was added a solution of lithium hexamethyldisilylamide in tetrahydrofuran (247 mL of 1.0 M; 1.1 eq.) via syringe under nitrogen atmosphere. The resulting yellow solution was allowed to stir at 0°C for 1.5 hours, then a solution of 3-cyclopentoxo-4-methoxybenzaldehyde (49.4 g; 225 mmol) in dry tetrahydrofuran (150 mL) was added dropwise via addition funnel over 0.5 hour. The resulting orange solution

was allowed to stir at 0°C for 2 hours, then was warmed to room temperature and stirred overnight. The reaction then was quenched with the addition of water (400 mL) and extracted with ether (2 x 300 mL). The combined organic layers were washed with 1N aqueous hydrochloric acid (250 mL), saturated aqueous bicarbonate (250 mL), and brine (250 mL), then dried over MgSO₄, filtered and concentrated in vacuo to provide the unsaturated ester as a brown liquid (68.4 g; 100%).

¹H NMR (CDCl₃, 400 MHz) δ 7.64 (s, 1H), 7.01-6.96 (c, 2H), 6.87 (m, 1H), 4.77 (m, 1H), 4.26 (q, 2H). LRMS (Electrospray, positive): Da/e 305.3 (m+1).

A suspension of (E)-3-(3-cyclopentoxymethyl-4-methoxyphenyl)-2-methyl-acrylic acid ethyl ester (30 g, 98.6 mmol) and LiOH•H₂O (lithium hydroxide hydrate) (5.0 g, 119.2 mmol, 1.2 equiv.) in methanol-water (4:1, 100 mL) was stirred at room temperature for 24 hours. Methanol was removed under reduced pressure and the resulting residue was dissolved in water (100 mL), washed with three 100 mL portions of ethyl acetate, neutralized with 1.0 N HCl (hydrochloric acid) (100 mL), and extracted with two 150 mL portions of ethyl acetate. The extract was washed with brine, dried over anhydrous Na₂SO₄ (sodium sulfate), and concentrated under reduced pressure to afford the desired product as a light yellow powder (18.2 g, 66% yield). ¹H NMR (300 MHz, CDCl₃) δ (ppm): 12.4 (br. s, 1H, COOH), 7.56 (s, 1H, olefinic), 7.04 (m, 3H, aromatic), 4.81 (m, 1H), 3.78 (s, 3H, OCH₃), 2.06 (s, 3H, CH₃), 1.91-1.57 (m, 8H, cyclopentyl).

Intermediate A was prepared in an alternative method as follows:

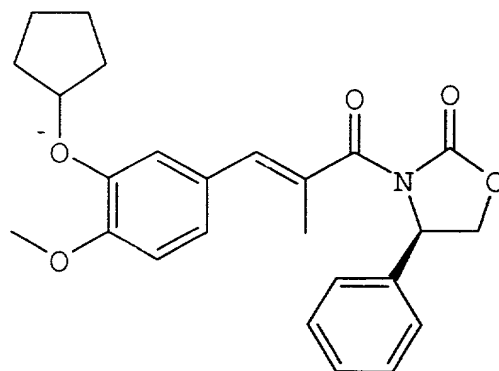
To a stirred solution of the ethyl ester (68.4 g; 225 mmol) in dioxane (400 mL) was added a solution of lithium hydroxide monohydrate (14.0 g; 332 mmol; 1.5 eq.) in water (200 mL) at room temperature and under a nitrogen atmosphere. A slight exotherm was observed. The resulting cloudy yellow solution was heated to 80°C (oil bath) for 1.5 hours. After heating for 0.5 hour, the reaction became clear, but required an additional 1.5 hours to complete the reaction, as evaluated by TLC. The resulting solution was allowed to cool to room temperature, diluted with ether (500 mL), then was washed with 1 M aqueous phosphoric acid (H₃PO₄). This aqueous layer then was extracted with ethyl acetate (2 x 200 mL) and the combined ethyl acetate and ether layers were washed with brine (250 mL), dried over MgSO₄, filtered and concentrated *in vacuo* to provide Intermediate A as an orange solid (55 g; 88%).

¹H NMR (CDCl₃, 400 MHz): δ 7.76 (s, 1H), 7.06-7.00 (c, 2H), 6.89 (m, 1H), 4.78 (m, 1H), 3.88 (s, 3H), 2.17 (s, 3H), 1.97-1.83 (c, 6H), 1.64-1.61 (c, 2H).

LRMS (Electrospray, negative): Da/e 275.3 (M-1).

Intermediate B

Preparation of 3-[(2E)-3-(3-cyclopentyloxy-4-methoxyphenyl)-2-methylprop-2-enoyl]-(4R)-4-phenyl-1,3-oxazolidin-2-one



To a cooled (0°C), stirred slurry of Intermediate A (55 g; 199 mmol) in anhydrous dichloromethane (400 mL) was added a solution of oxalyl chloride in dichloromethane (109 mL of 2.0 M; 218 mmol; 1.1 eq.) via a syringe under a calcium chloride-dried atmosphere over 10 minutes. Vigorous bubbling was observed. The resulting dark solution was allowed to stir at 0°C for 15 minutes, then a catalytic amount of dimethylformamide was added via syringe (0.3 mL). The resulting solution was allowed to continue stirring at 0°C for 0.5 hour as the bubbling subsided, and then was allowed to warm to room temperature and stirred overnight (17 hours). The reaction was diluted with ethyl acetate (500 mL) and was carefully quenched with water (250 mL). After vigorously stirring this mixture for 1 hour, the layers were separated, and the organic layer was washed with additional water (400 mL) and brine (400 mL), then dried over MgSO₄, filtered and

concentrated in vacuo to provide an acid chloride as a brown solid (57.5 g; 98%).

¹H NMR (CDCl₃, 400 MHz): δ 7.98 (s, 1H), 7.11-7.02 (c, 2H), 6.92 (m, 1H), 4.79 (m, 1H), 3.90 (s, 3H),
5 2.22 (s, 3H), 2.01-1.82 (c, 6H), 1.68-1.62 (c, 2H).

To a cooled (-78°C), mechanically stirred solution of R-phenyl oxazolidinone (10.0 g; 61.3 mmol) in dry tetrahydrofuran (400 mL) was added a solution of n-butyllithium in hexanes (27 mL of 2.5
10 M; 1.1 eq.) via syringe under nitrogen atmosphere. The resulting solution was allowed to stir at -78°C for 0.8 hour, then a solution of the acid chloride (19.9 g; 67.4 mmol; 1.1 eq.) in tetrahydrofuran (100 mL) was added via cannulae. After stirring at -78°C
15 for 15 minutes, the reaction mixture was allowed to slowly warm to 0°C over 40 minutes, during which time the reaction became a thick slurry. After stirring at 0°C for 2.5 hours, the reaction was quenched with saturated, aqueous ammonium chloride
20 (300 mL), and the bulk of the tetrahydrofuran was removed at reduced pressure. The residue then was extracted with chloroform (3 x 700 mL) and the combined organic layers were washed with water (300 mL) and brine (300 mL), then dried over MgSO₄, filtered
25 and concentrated in vacuo to provide about 33 g of a light orange solid. This material was suspended in 10% ethyl acetate in hexane (1.2 L), and vigorously stirred overnight. The resulting fine powdery solids were collected on a Buchner funnel with suction
30 and then dried in vacuo to provide Intermediate B as a tan powder (21.8 g; 88%).

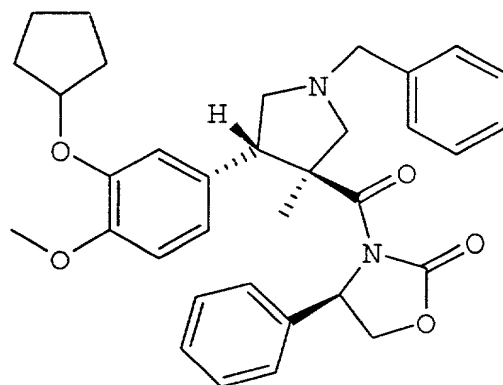
¹H NMR (CDCl₃, 400 MHz): δ 7.41-7.37 (c, 5H), 7.06 (s, 1H), 7.01-6.97 (c, 2H), 6.86 (m, 1H), 5.54 (t,

1H), 4.77-4.73 (c, 2H), 4.29 (t, 1H), 3.87 (s, 3H),
2.17 (s, 3H), 1.97-1.82 (c, 6H), 1.62-1.56 (c, 2H).

Similarly, the enantiomer of Intermediate
B can be prepared using S-(-)-4-phenyl oxazolidi-
5 none.

Intermediate C

Preparation of (4R)-3-{[(3S,4S)-4-(3-cyclopentyloxy-
4-methoxyphenyl)-3-methyl-1-benzylpyrrolidin-3-yl]
10 carbonyl)-4-phenyl-1,3-oxazolidin-2-one



To a cooled (-4°C), stirred slurry of
Intermediate B (9.30 g; 22.8 mmol) and N-(methoxy-
25 methyl)-N-(trimethylsilylmethyl)benzylamine (11.7
mL; 45.6 mmol; 2 eq.) in chloroform (65 mL) was
added a solution of trifluoroacetic acid in chloro-
form (4.6 mL of 1.0 M; 4.6 mmol; 0.2 eq.) via sy-
ringe under nitrogen atmosphere. The resulting
30 slurry was allowed to stir at about 0°C for 4 hours,
and then at about 15°C overnight (water bath). The
resulting cloudy solution then was recooled to -4°C
and treated with additional N-(methoxymethyl)-N-
(trimethylsilylmethyl)benzylamine (5.9 mL; 22.8

mmol; 1 eq.) via syringe, and allowed to stir for 5 hours during which time the reaction became homogeneous. TLC (5% Et₂O in CH₂Cl₂) show the reaction was complete. The bulk of the chloroform was removed at reduced pressure and the residue was diluted with ethyl acetate (250 mL), then washed successively with 1 N aqueous hydrochloric acid (2 x 50 mL), 1 N aqueous sodium hydroxide (50 mL), and brine (50 mL). The organic layer then was dried over MgSO₄, filtered, and concentrated in vacuo to give an orange semisolid (13.9 g). Purification via flash chromatography on silica gel (2% ether in dichloromethane) provided the major diastereomer pyrrolidine as a white foam (8.25 g; 65%).

Diastereomeric selectivity about 10:1 (HPLC).

¹H NMR (CDCl₃, 400 MHz): δ 7.42-7.21 (c, 10H), 6.95 (s, 1H), 6.81 (s, 2H), 5.55 (dd, 1H), 4.74 (t, 1H), 4.68 (m, 1H), 4.10 (dd, 1H), 3.93 (t, 1H), 3.70 (d, 1H), 3.68 (s, 3H), 3.56 (d, 1H), 3.42 (d, 1H), 2.72 (m, 2H), 2.64 (d, 1H), 2.48 (m, 1H), 1.85-1.78 (c, 2H), 1.75-1.61 (c, 4H), 1.57-1.53 (c, 2H), 0.96 (s, 3H). LRMS (Electrospray, positive): Da/e 555.2 (m+1).

25 Intermediate D

Preparation of (3S,4S)-4-(3-cyclopentyloxy-4-methoxyphenyl)-3-methyl-1-benzylpyrrolidine-3-carbaldehyde

30

Reduction/Oxidation Procedure:

To a cooled (-78°C), stirred solution of Intermediate C (15.09 g; 27.2 mmol) in toluene (250mL) was added a solution of lithium aluminum

hydride in tetrahydrofuran (16.3 mL; 1.0 M; 16.3 mmol; 0.6 eq.) via syringe under nitrogen atmosphere. Vigorous bubbling was observed. The resulting solution was allowed to stir at -78°C for 2
5 hours, then the cooling bath was removed and quenched with the successive addition of water (0.62 mL), 15% aqueous sodium hydroxide (0.62 mL) and more water (1.9 mL). The resulting mixture was allowed to warm to room temperature, stirred for 30 minutes,
10 and then was diluted with ether (500 mL) and dried over MgSO₄. Filtration and concentration *in vacuo* provided the alcohol (with some aldehyde present) as a semisolid (14.8 g). This material was used immediately without further purification.

15 To a cooled (-78°C), stirred solution of oxalyl chloride in dichloromethane (10.9 mL; 2.0 M; 21.8 mmol; 0.8 eq.) in more dichloromethane (75 mL) was added dimethylsulfoxide (3.1 mL; 43.5 mmol; 1.6 eq.) via syringe under nitrogen atmosphere. Vigor-
20 ous bubbling was observed. After stirring at -78°C for 20 minutes, a solution of the crude alcohol in dichloromethane (75 mL) was added via cannulae. The resulting yellow solution was allowed to stir at -78°C for 20 minutes, then triethylamine (15.2 mL;
25 109 mmol; 4 eq.) was added via syringe. The reaction was allowed to stir at -78°C for 20 minutes, then warm to room temperature and stirred for an additional 1 hour. The reaction was quenched with the addition of brine (150 mL), then extracted with
30 dichloromethane (2 x 100 mL). The combined organic fractions were dried over MgSO₄, filtered, then concentrated *in vacuo* to provide the crude aldehyde. Purification by flash silica gel chromatography (25%

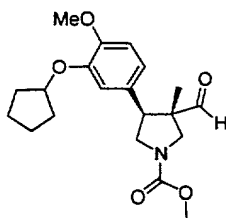
ethyl acetate in hexanes) provided Intermediate D as a clear, colorless oil (9.8 g; 92%).

^1H NMR (CDCl_3 , 400 MHz): δ 9.64 (s, 1H), 7.37-7.26 (c, 5H), 6.78-6.76 (c, 2H), 6.70 (m, 1H), 4.74 (m, 1H), 3.82 (s, 3H), 3.70 (m, 1H), 3.64-3.62 (c, 2H), 3.18-3.13 (c, 2H), 2.84 (t, 1H), 2.41 (d, 1H), 1.94-1.83 (c, 6H), 1.63-1.59 (c, 2H), 0.74 (s, 3H).

LRMS (Electrospray, positive): Da/e 394.3 (m+1).

10 Intermediate 1

Preparation of 4-(S)-(3-Cyclopentyloxy-4-methoxy-phenyl)-3-(S)-formyl-3-methylpyrrolidine-1-carboxylic acid methyl ester



Intermediate D was dissolved in acetonitrile (4.8 mL) and treated with methyl chloroformate (176 μL , 2.3 mmol). The resulting solution was heated to reflux for 4 hours, and thin layer chromatography indicated complete consumption of Intermediate D. The reaction mixture then was concentrated in vacuo and purified directly by column chromatography on SiO_2 using hexanes/ethyl acetate (3:1) as eluant. Intermediate 1 was recovered as a light yellow oil (110 mg).

^1H NMR (CDCl_3 , 400 MHz) δ : 9.62 (s, 1H), 6.83-6.79 (d, 1H), 6.65 (dd, 1H), 6.61 (d, 1H), 4.74-4.70 (brd s, 1H), 3.94-3.64 (m, 3H), 3.83 (s, 3H), 3.76 (s,

3H), 3.62-3.55 (m, 1H), 3.38-3.28 (dd, 1H), 1.95-1.75 (m, 6H), 1.66-1.57 (m, 2H), 0.91 (s, 3H).

The following illustrates the synthesis of various intermediates and compounds of structural formula (II). The following examples are provided for illustration and should not be construed as limiting.

In the examples, the following abbreviations are used: NaOH (sodium hydroxide), CH₂Cl₂ (dichloromethane), Na₂SO₄ (sodium sulfate), EtOAc (ethyl acetate), Ph (phenyl), MeOH (methanol), K₂CO₃ (potassium carbonate), μ mol (micromole), atm (atmosphere), MgSO₄ (magnesium sulfate), LiOH (lithium hydroxide), N (normal), mmol (millimole), mL (milliliter), and H₃PO₄ (phosphoric acid).

The following general procedures were used to prepare Examples 1-28.

Reductive Amination Procedure:

To a stirred solution of aldehyde (0.24 mmol) in dichloroethane (1 mL) was added benzylamine (0.24 mmol) followed by sodium triacetoxyborohydride (0.34 mmol) at room temperature under a nitrogen atmosphere. After stirring for 5 hours, the reaction was diluted with 1 N aqueous NaOH (0.3 mL), and CH₂Cl₂ (10 mL). The layers were separated and the aqueous layer extracted with another 5 mL of CH₂Cl₂. Combined organic layers then were washed with water and brine, dried (Na₂SO₄), filtered, and concentrated *in vacuo*. The residue was purified via flash chro-

matography (3:2 EtOAc:hexanes on silica gel) to provide the product.

Hydrogenation Procedure:

5

To a stirred solution of the starting material (75 μ mol) in 95% ethanol (1 mL) was treated with Pearlman's catalyst (palladium hydroxide on carbon, 10 mg), then placed under a hydrogen atmosphere (1 atm). After stirring at room temperature for 24 hours, the reaction was filtered through GF/F filter paper with suction on a Buchner funnel and washed with 20 mL of 95% ethanol. Concentration of the filtrate provided the product.

15

Sulfonylation Procedure:

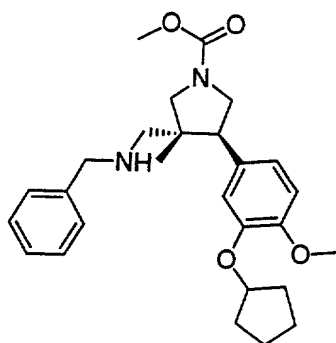
To a stirred solution of starting material (0.1 mmol) in 1,4-dioxane (0.3 mL) was added, successively, aqueous K_2CO_3 (0.6 mL of 0.65 M; 4 eq) and a solution of the sulfonyl chloride (0.12 mmol) in 1,4-dioxane (0.3 mL) at room temperature. The resulting solution was allowed to stir at room temperature for 2 hours. The reaction was diluted with 1:1 hexanes:EtOAc (30 mL) and washed successively with water (20 mL) and brine (20 mL), then dried ($MgSO_4$), filtered, and concentrated *in vacuo* to provide the sulfonamide.

30 **Hunig's Base Acylation Procedure:**

To a stirred solution of starting material (0.14 mmol) and Hunig's base (0.20 mmol) in dry

CH₂Cl₂ (1 mL) was added the acid chloride (0.14 mmol) via syringe at room temperature under a nitrogen atmosphere. After stirring for 1 hour, the reaction was diluted with CH₂Cl₂ (30 mL) and washed successively with 1 N aqueous HCl (2 x 10 mL) and brine (10 mL), then dried (Na₂SO₄), filtered, and concentrated in vacuo. The residue was purified via radial chromatography (1 mm chromatotron plate with 3% MeOH in CH₂Cl₂) to provide the amide.

Example 1



R=CH₂Ph

Methyl (4S,3R)-4-(3-cyclopentyloxy-4-methoxyphenyl)-3-methyl-3-{[benzylamino]methyl}pyrrolidine carboxylate

Prepared by the reductive amination procedure.

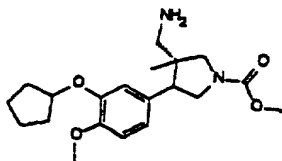
¹H NMR (CDCl₃, 400 MHz; mixture of rotomers) δ:

7.34-7.28 (m, 5H), 6.77 (d, 1H), 6.67-6.64 (m, 2H), 4.70 (c, 1H), 3.84-3.65 (m, 9H), 3.45-3.24 (m, 4H), 2.54 (q, 2H), 1.86-1.79 (m, 6H), 1.62-1.55 (m, 2H), 0.78 (d, 3H).

LRMS (Electrospray, positive): Da/e 453.2 (m+1).

Example 2

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R=H

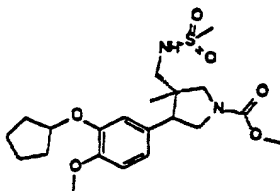
15 Methyl (4S,3R)-3-(aminomethyl)-4-(3-cyclopentyloxy-4-methoxyphenyl)-3-methylpyrrolidinecarboxylate

Prepared via the hydrogenation procedure.

¹H NMR (CDCl₃, 400 MHz; mixture of rotomers) δ: 6.80 (d, 1H), 6.71-6.65 (m, 2H), 4.73 (c, 1H), 3.86-3.68 (m, 7H), 3.43-3.17 (m, 4H), 2.65 (br s, 2H), 1.97-1.80 (m, 6H), 1.69-1.56 (m, 2H), 0.77 (d, 3H).
20 LRMS (Electrospray, positive): Da/e 363.3 (m+1).

Example 3

5



10

R=SO₂CH₃

Methyl (3S,4S)-4-(3-cyclopentyloxy-4-methoxyphenyl)-
3-methyl-3-([methylsulfonyl]amino)methylpyrrolidine-
1-carboxylate

15

Prepared via the sulfonylation procedure.

¹H NMR (CDCl₃, 400 MHz; mixture of rotomers) δ: 6.80
(d, 1H), 6.69-6.67 (m, 2H), 4.81-4.72 (m, 1.5H),
4.61 (t, 0.5H), 3.88-3.66 (m, 8H), 3.44-3.35 (m,
20 2H), 3.20 (t, 0.5H), 3.14-3.05 (m, 2.5H), 2.95 (s,
3H), 1.91-1.80 (m, 6H), 1.63-1.56 (m, 2H), 0.82 (d,
3H).

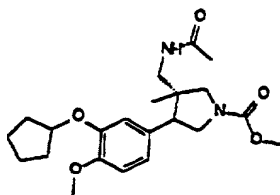
LRMS (Electrospray, positive): Da/e 441.2 (m+1).

LRMS (Electrospray, negative): Da/e 439.2 (m-1).

25

Example 4

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R=COCH₃,

Methyl (4S,3R)-3-[(acetylamino)methyl]-4-(3-cyclopentyloxy-4-methoxyphenyl)-3-methylpyrrolidine-carboxylate

15

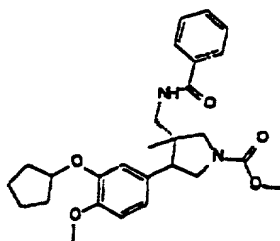
Prepared via the Hunig's base mediated acylation procedure.

¹H NMR (CDCl₃, 400 MHz; mixture of rotomers) δ: 6.80 (d, 1H), 6.69-6.67 (m, 2H), 5.58 (br t, 0.5H), 5.52 (br t, 0.5H), 4.74 (c, 1H), 3.86-3.64 (m, 7H), 3.39-3.19 (m, 5H), 3.08 (t, 1H), 1.94 (s, 3H), 1.91-1.79 (m, 6H), 1.63-1.58 (m, 2H), 0.80 (d, 3H).

LRMS (Electrospray, positive): Da/e 405.3 (m+1).

25

Example 5



R=COPh

Methyl (4S,3R)-4-(3-cyclopentyloxy-4-methoxyphenyl)-
3-methyl-3-[(phenylcarbonylamino)methyl]pyrroli-
dinecarboxylate

Prepared via the Hunig's base mediated acylation
procedure.

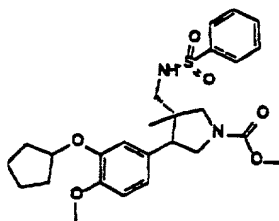
¹H NMR (CDCl₃, 400 MHz; mixture of rotomers) δ:

7.55-7.45 (m, 3H), 7.37 (t, 2H), 6.83 (d, 1H), 6.77-
6.74 (m, 2H), 6.14 (br s, 1H), 4.73 (c, 1H), 3.90-
3.59 (m, 9H), 3.50-3.30 (m, 3H), 3.18 (t, 1H), 1.97-
1.74 (m, 6H), 1.64-1.55 (m, 2H), 0.94 (d, 3H).

LRMS (Electrospray, positive): Da/e 467.3 (m+1).

Example 6

5



10

R=SO₂Ph

15 Methyl (3S,4S)-4-(3-cyclopentyloxy-4-methoxyphenyl)-
3-methyl-3-{[phenylsulfonyl]amino}methylpyrrolidine-
dinecarboxylate

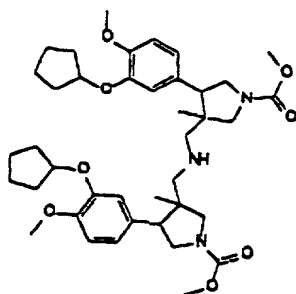
Prepared via the sulfonylation procedure.

¹H NMR (CDCl₃, 400 MHz; mixture of rotomers) δ: 7.84
20 (dd, 2H), 7.61-7.50 (m, 3H), 6.77 (dd, 1H), 6.64-
6.60 (m, 2H), 5.01 (t, 0.5H), 4.93 (t, 0.5H), 4.72
(c, 1H), 3.95-3.59 (m, 9H), 3.32 (c, 1H), 3.19 (t,
0.5H), 3.07 (t, 0.5H), 2.93-2.82 (m, 2H), 1.97-1.74
(m, 6H), 1.64-1.53 (m, 2H), 0.75 (s, 3H).

25 LRMS (Electrospray, positive): Da/e 503.2 (m+1).

LRMS (Electrospray, negative): Da/e 501.2 (m-1).

Example 7



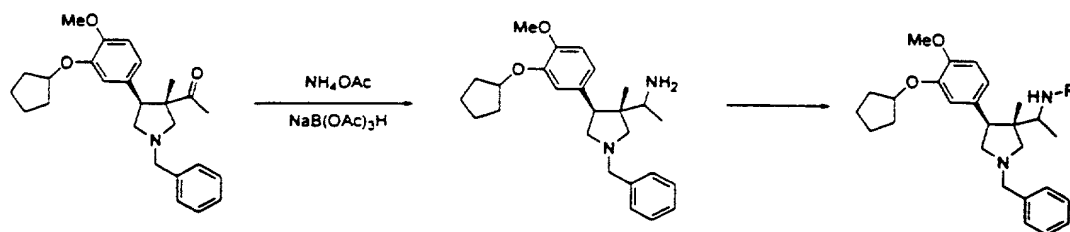
R=DIMER

Bis{[(4S,3R)-4-(3-cyclopentyloxy-4-methoxyphenyl)-3-methyl-1-carboxymethylpyrrolidin-3-yl]methyl}amine

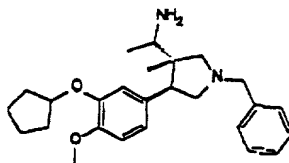
Prepared via the reductive amination procedure with ammonium acetate.

^1H NMR (CDCl_3 , 400 MHz; mixture of rotomers) δ : 6.80 (dd, 2H), 6.72-6.64 (m, 4H), 4.74 (m, 2H), 3.93-3.67 (m, 16H), 3.56-3.23 (m, 8H), 2.51 (q, 1H), 2.06-1.81 (m, 2H), 1.70-1.53 (m, 4H), d, 3H).

LRMS (Electrospray, positive): Da/e 708.1 (m+1).



Example 8



R=H

1-[(3S,4S)-4-(3-Cyclopentyloxy-4-methoxyphenyl)-3-methyl-1-benzylpyrrolidin-3-yl]ethanamine Less Polar Diastereomer

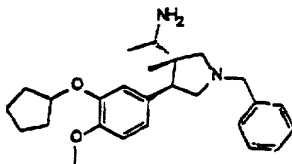
Prepared via the reductive amination procedure.

¹H NMR (CDCl₃, 400 MHz) δ : 7.37-7.21 (c, 5H), 6.78 (d, 1H), 6.69-6.67 (c, 2H), 4.75 (m, 1H), 3.82 (s, 3H), 3.56 (s, 2H), 2.86 (m, 1H), 2.73 (m, 2H), 2.50 (t, 1H), 2.01 (t, 1H), 1.93-1.78 (c, 7H), 1.67-1.56 (c, 2H), 1.28 (br s, 2H), 0.89 (d, 3H), 0.86 (s, 3H).

LRMS (Electrospray, positive): Da/e 409.3 (m+1).

Example 9

5



10

R=H

1-[(3S,4S)-4-(3-Cyclopentyloxy-4-methoxyphenyl)-3-methyl-1-benzylpyrrolidin-3-yl]ethan-1-amine More Polar

15 Diastereomer

Prepared via the reductive amination procedure.

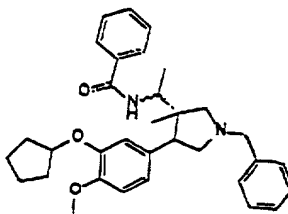
¹H NMR (CDCl₃, 400 MHz) δ: 7.39-7.22 (c, 5H), 6.93 (s, 1H), 6.78-6.69 (c, 2H), 4.75 (m, 1H), 3.82 (s, 3H), 3.68 (d, 1H), 3.56 (d, 1H), 3.17 (t, 1H), 2.92-2.81 (c, 3H), 2.62 (d, 1H), 2.37 (d, 1H), 1.98-1.81 (c, 6H), 1.67-1.57 (c, 2H), 1.05 (d, 3H), 0.63 (s, 3H).

LRMS (Electrospray, positive): Da/e 409.3 (m+1).

25

005120-4942004

Example 10



R=COPh

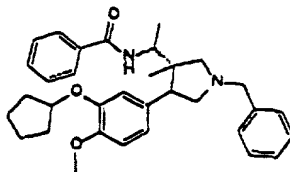
N-{1-[(3S,4S)-4-(3-Cyclopentyloxy-4-methoxyphenyl)-
3-methyl-1-benzylpyrrolidin-3-yl]ethyl}benzamide
Less Polar Diastereomer

Prepared via the Hunig's base mediated acylation
procedure.

¹H NMR (CDCl₃, 400 MHz) δ: 8.47 (d, 1H), 7.81 (d, 2H), 7.51 (t, 1H), 7.42 (t, 2H), 7.38-7.29 (m, 5H), 6.77 (t, 1H), 6.73 (d, 2H), 4.78 (c, 1H), 4.01 (c, 1H), 3.82 (s, 3H), 3.72 (d, 1H), 3.62 (d, 1H), 3.40-3.26 (m, 3H), 2.58 (t, 1H), 2.13 (d, 1H), 1.96-1.74 (m, 6H), 1.70-1.53 (m, 2H), 1.22 (d, 3H), 0.60 (s, 3H).

LRMS (Electrospray, positive): Da/e 513.3 (m+1).

Example 11



10 R=COPh

N-{1-[(3S,4S)-4-(3-Cyclopentyloxy-4-methoxyphenyl)-
3-methyl-1-benzylpyrrolidin-3-yl]ethyl}benzamide

More Polar Diastereomer

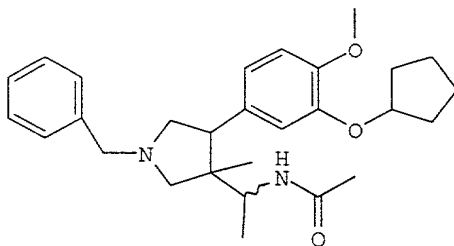
15 Prepared via the Hunig's base mediated acylation
procedure.

¹H NMR (CDCl₃, 400 MHz) δ : 7.92 (br s, 1H), 7.81 (d, 2H), 7.53 (t, 1H), 7.46 (t, 2H), 7.20-7.13 (m, 5H), 6.78-6.72 (m, 3H), 4.64 (t, 1H), 4.00 (c, 1H), 3.84-
20 3.77 (m, 4H), 3.57 (q, 2H), 3.43 (t, 1H), 3.25 (t, 1H), 2.81 (d, 1H), 2.65 (t, 1H), 2.34 (d, 1H), 1.87-1.79 (m, 6H), 1.65-1.52 (m, 2H), 1.29 (d, 3H), 0.80 (s, 3H).

LRMS (Electrospray, positive): Da/e 513.3 (m+1).

25

Example 12



R=COCH₃

N-{1-[(3S,4S)-4-(3-Cyclopentyloxy-4-methoxyphenyl)-
3-methyl-1-benzylpyrrolidin-3-yl]ethyl}acetamide

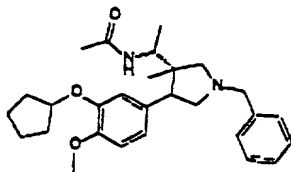
15 Less Polar Diastereomer

Prepared via the Hunig's base mediated acylation
procedure.

¹H NMR (CDCl₃, 400 MHz) δ: 7.35-7.24 (m, 5H), 6.87
20 (br s, 1H), 6.82 (d, 1H), 6.75 (d, 1H), 6.71 (dd,
1H), 4.74 (c, 1H), 3.85-3.80 (m, 4H), 3.68 (d, 1H),
3.49 (d, 1H), 3.26 (t, 1H), 3.12 (t, 1H), 2.66 (t,
1H), 2.31 (d, 1H), 1.96 (s, 3H), 1.95-1.80 (m, 6H),
1.64-1.58 (m, 2H), 1.15 (d, 3H), 0.72 (s, 3H).

25 LRMS (Electrospray, positive): Da/e 451.3 (m+1).

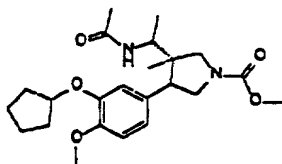
Example 13



- 10 R=COCH₃
N-{1-[(3S,4S)-4-(3-Cyclopentyloxy-4-methoxyphenyl)-
3-methyl-1-benzylpyrrolidin-3-yl]ethyl}acetamide
More Polar Diastereomer
- 15 Prepared via the Hunig's base mediated acylation
procedure.
- ¹H NMR (CDCl₃, 400 MHz) δ: 7.61 (br d, 1H), 7.36-
7.26 (m, 5H), 6.78-6.70 (m, 3H), 4.77 (c, 1H), 3.86-
3.78 (m, 5H), 3.63 (d, 1H), 3.27 (t, 1H), 3.22 (t,
20 1H), 3.06 (d, 1H), 2.58 (t, 1H), 2.03 (s, 3H), 1.96-
1.80 (m, 6H), 1.62-1.57 (m, 2H), 1.09 (d, 3H), 0.53
(s, 3H).
- LRMS (Electrospray, positive): Da/e 451.3 (m+1).

Example 14

5



10 R=COCH₃

3-(S)-(1-Acetylaminoethyl)-4-(S)-(3-cyclopentyloxy-
4-methoxyphenyl)-3-methylpyrrolidine-1-carboxic acid
methyl ester

Less Polar Diastereomer

15

Prepared via the Hunig's base mediated acylation
procedure.

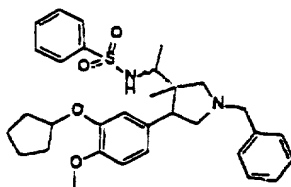
¹H NMR (CDCl₃, 400 MHz) δ: 7.61 (br d, 1H), 7.38-
7.26 (m, 5H), 6.78-6.70 (m, 3H), 4.77 (c, 1H), 3.86-
20 3.76 (m, 4H), 3.63 (d, 2H), 3.27 (t, 1H), 3.22 (t,
1H), 3.06 (d, 1H), 2.58 (t, 1H), 2.06 (d, 1H), 2.03
(s, 3H), 1.96-1.78 (m, 6H), 1.65-1.55 (m, 2H), 1.09
(d, 3H), 0.53 (s, 3H).

LRMS (Electrospray, positive): Da/e 451.3 (m+1).

25

Example 15

5



10 R=SO₂Ph

{1-[(3S,4S)-4-(3-Cyclopentyloxy-4-methoxyphenyl)-3-methyl-1-benzylpyrrolidin-3-yl]ethyl}-(phenylsulfonyl)amine

More Polar Diastereomer

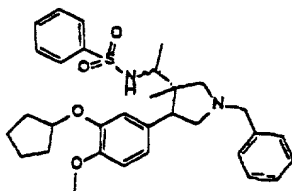
15

Prepared via the Hunig's base mediated acylation procedure.

¹H NMR (CDCl₃, 400 MHz) δ: 7.89 (d, 2H), 7.54-7.41 (m, 2H), 7.40-7.34 (m, 5H), 7.31 (c, 1H), 6.66 (dd, 1H), 6.34 (s, 1H), 6.24 (d, 1H), 4.67 (c, 1H), 3.78 (s, 3H), 3.66 (d, 1H), 3.57 (d, 1H), 3.14-3.06 (m, 3H), 2.69 (t, 1H), 2.39 (t, 1H), 1.96 (d, 1H), 1.91-1.77 (m, 6H), 1.69-1.55 (m, 2H), 1.11 (d, 3H), 0.40 (s, 3H).

25 LRMS (Electrospray, positive): Da/e 549.2 (m+1).

Example 16



R=SO₂Ph

{1-[(3S,4S)-4-(3-cyclopentyloxy-4-methoxyphenyl)-3-methyl-1-benzylpyrrolidin-3-yl]ethyl}-
(phenylsulfonyl)amine

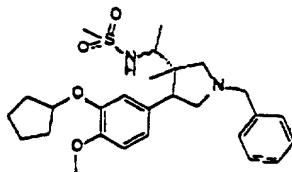
Less Polar Diastereomer

Prepared via the Hunig's base mediated acylation procedure.

¹H NMR (CDCl₃, 400 MHz) δ: 7.64 (d, 2H), 7.52 (t, 1H),
7.43-7.31 (m, 7H), 6.77-6.66 (m, 3H), 4.73 (c, 1H),
3.80 (s, 3H), 3.53 (d, 2H), 3.47 (t, 1H), 3.29 (t,
1H), 2.87 (q, 1H), 2.44 (t, 1H), 2.29 (d, 1H), 2.11
(d, 1H), 1.93-1.77 (m, 6H), 1.61-1.56 (m, 2H) 1.21
(d, 3H), 0.54 (s, 3H).

LRMS (Electrospray, positive): Da/e 549.2 (m+1).

Example 17



10

R=SO₂CH₃

{1-[(3S,4S)-4-(3-Cyclopentyloxy-4-methoxyphenyl)-3-methyl-1-benzylpyrrolidin-3-yl]ethyl}-(methylsulfonyl)amine

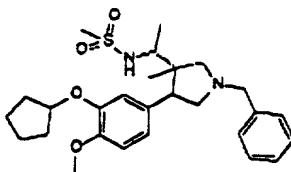
15 More Polar Diastereomer

Prepared via the Hunig's base mediated acylation procedure.

¹H NMR (CDCl₃, 400 MHz) δ: 7.37-7.28 (m, 5H), 6.78-6.71 (m, 3H), 4.76 (c, 1H), 3.81 (s, 3H), 3.70 (d, 1H), 3.54 (d, 1H), 3.50 (d, 1H), 3.32 (t, 1H), 3.01 (d, 1H), 2.98 (s, 3H), 2.56 (t, 1H), 2.02 (t, 1H), 1.98-1.83 (m, 6H), 1.62-1.53 (m, 2H), 1.18 (d, 3H), 0.52 (s, 3H).

25 LRMS (Electrospray, positive): Da/e 487.3 (m+1).

5


$$R = \text{SO}_2\text{CH}_3$$

15

Less Polar Diastereomer

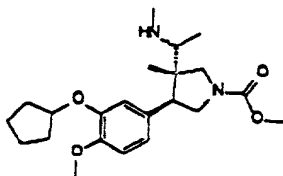
20

¹H NMR (CDCl₃, 400 MHz) δ: 7.40-7.27 (m, 5H), 6.80-6.75 (m, 3H), 4.75 (c, 1H), 3.87-3.78 (m, 4H), 3.47 (t, 1H), 3.38 (d, 1H), 3.36 (t, 1H), 3.22 (q, 1H), 2.78 (d, 1H), 2.63-2.57 (m, 4H), 2.26 (d, 1H), 1.95-1.79 (m, 1H), 1.70-1.58 (m, 2H), 1.28 (d, 3H), 0.68 (s, 3H).

25 LRMS (Electrospray, positive): Da/e 487.3 (m+1).

Example 19

5



10

R=CH₃

Methyl (3S,4S)-4-(3-cyclopentyloxy-4-methoxyphenyl)-
3-methyl-3-[(methyamino)ethyl]pyrrolidine carbox-
ylate

Mixture of Diastereomers

15

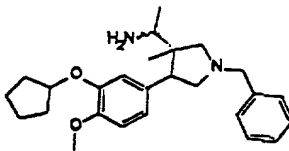
Prepared from the methyl ketone via the reductive
amination procedure with methylamine.

¹H NMR (CDCl₃, 400 MHz) δ: 6.79 (d, 1H), 6.68-6.65
(m, 2H), 4.71 (c, 1H), 3.96-3.62 (m, 11H), 3.37 (d,
20 0.5H), 3.27 (d, 0.5H), 2.15 (d, 3H), 2.01-1.74 (m,
6H), 1.62-1.55 (m, 2H), 1.01 (d, 3H).

LRMS (Electrospray, positive): Da/e 391.4 (m+1).

Example 20

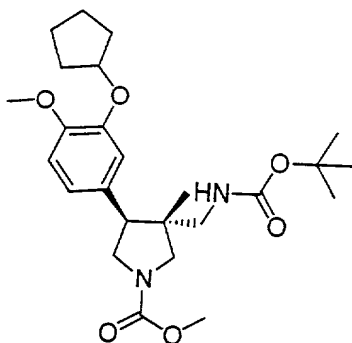
25



30

Example 21

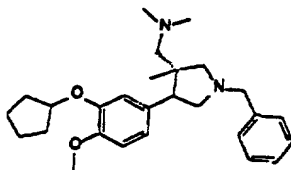
5



10

Example 22

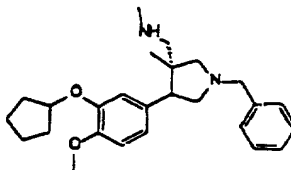
15



20

Example 23

25



30

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CCOC(=O)N1CC[C@H](C1)c2ccc(Oc3ccccc3)cc2

Example 25

COC(=O)N1CCC(C1c2ccc(OC3CC3)cc2)C4CC4

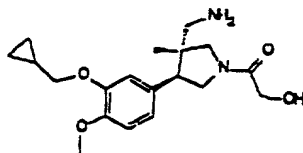
Example 26

CC(C)(C)C(=O)N1CC[C@H](C1)C2=CC(OC3CC3)C(OC4CC4)=CC=C2

30

Example 27

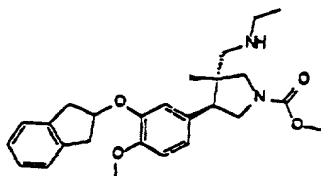
5



10

Example 28

15



20

The compounds of structural formula (II) were tested for an ability to inhibit PDE4. The ability of a compound to inhibit PDE4 activity is related to the IC_{50} value for the compound, i.e., the concentration of inhibitor required for 50% inhibition of enzyme activity. The IC_{50} value for compounds of structural formula (II) were determined using recombinant human PDE4.

The compounds of the present invention typically exhibit an IC_{50} value against recombinant human PDE4 of less than about 100 μM , and preferably less than about 50 μM , and more preferably less than about 25 μM . The compounds of the present invention

typically exhibit an IC_{50} value against recombinant human PDE4 of less than about 5 μM , and often less than about 1 μM . To achieve the full advantage of the present invention, a present PDE4 inhibitor has
5 an IC_{50} of about 1 nM to about 25 μM .

The IC_{50} values for the compounds were determined from concentration-response curves typically using concentrations ranging from 0.1 pM to 500 μM . Tests against other PDE enzymes using stan-
10 dard methodology, as described in Loughney et al., *J. Biol. Chem.*, 271, pp. 796-806 (1996), also showed that compounds of the present invention are highly selective for the cAMP-specific PDE4 enzyme.

The compounds of structural formula (II)
15 also were tested for an ability to reduce TNF α secretion in human peripheral blood lymphocytes. The ability to reduce TNF α secretion is related to the EC_{50} values (i.e., the effective concentration of the compound capable of inhibiting 50% of the total
20 TNF α).

The compounds of the present invention typically exhibit an EC_{50} value of less than about 50 μM , and preferably less than about 25 μM , and more preferably less than about 15 μM . The compounds of
25 the present invention preferably exhibit a PBL/TNF α EC_{50} value of less than about 5 μM , and often less than about 0.10 μM . To achieve the full advantage of the present invention, a present PDE4 inhibitor has an EC_{50} value of about 10 nM to about 20 μM .

30 The production of recombinant human PDEs and the IC_{50} and EC_{50} determinations can be accomplished by well-known methods in the art. Exemplary methods are described as follows:

EXPRESSION OF HUMAN PDEs

Expression in Baculovirus-Infected Spodoptera fugiperda (Sf9) Cells

5

10 Baculovirus transfer plasmids were constructed using either pBlueBacIII (Invitrogen) or pFastBac (BRL-Gibco). The structure of all plasmids was verified by sequencing across the vector junctions and by fully sequencing all regions generated by PCR. Plasmid pBB-PDE1A3/6 contained the complete open reading frame of PDE1A3 (Loughney et al., J. Biol. Chem., 271, pp. 796-806 (1996)) in pBlue-
15 BacIII. Plasmid Hcam3aBB contained the complete open reading frame of PDE1C3 (Loughney et al. (1996)) in pBlueBacIII. Plasmid pBB-PDE3A contained the complete open reading frame of PDE3A (Meacci et al., Proc. Natl. Acad. Sci., USA, 89, pp. 3721-3725
20 (1992)) in pBlueBacIII.

Recombinant virus stocks were produced using either the MaxBac system (Invitrogen) or the FastBac system (Gibco-BRL) according to the manufacturer's protocols. In both cases, expression of
25 recombinant human PDEs in the resultant viruses was driven off the viral polyhedron promoter. When using the MaxBac[®] system, virus was plaque purified twice in order to insure that no wild type (occ+) virus contaminated the preparation. Protein expression was carried out as follows. Sf9 cells were
30 grown at 27°C in Grace's Insect culture medium (Gibco-BRL) supplemented with 10% fetal bovine serum, 0.33% TC yeastolate, 0.33% lactalbumin hydrolysate, 4.2 mM NaHCO₃, 10 µg/mL gentamycin, 100

units/mL penicillin, and 100 µg/mL streptomycin. Exponentially growing cells were infected at a multiplicity of approximately 2 to 3 virus particles per cell and incubated for 48 hours. Cells were
5 collected by centrifugation, washed with nonsupplemented Grace's medium, and quick-frozen for storage.

Expression in *Saccharomyces cerevisiae* (Yeast)

10
Recombinant production of human PDE1B, PDE2, PDE4A, PDE4B, PDE4C, PDE4D, PDE5, and PDE7 was carried out similarly to that described in Example 7 of U.S. Patent No. 5,702,936, incorporated herein by
15 reference, except that the yeast transformation vector employed, which is derived from the basic ADH2 plasmid described in Price et al., *Methods in Enzymology*, 185, pp. 308-318 (1990), incorporated
20 *Saccharomyces cerevisiae* host was the protease-deficient strain BJ2-54 deposited on August 31, 1998 with the American Type Culture Collection, Manassas, Virginia, under accession number ATCC 74465. Transformed host cells were grown in 2X SC-leu medium, pH
25 6.2, with trace metals, and vitamins. After 24 hours, YEP medium-containing glycerol was added to a final concentration of 2X YET/3% glycerol. Approximately 24 hr later, cells were harvested, washed, and stored at -70°C.

30

HUMAN PHOSPHODIESTERASE PREPARATIONS

Phosphodiesterase Activity Determinations

5 Phosphodiesterase activity of the prepara-
tions was determined as follows. PDE assays utiliz-
ing a charcoal separation technique were performed
essentially as described in Loughney et al. (1996).
In this assay, PDE activity converts [32P]cAMP or
10 [32P]cGMP to the corresponding [32P]5'-AMP or
[32P]5'-GMP in proportion to the amount of PDE ac-
tivity present. The [32P]5'-AMP or [32P]5'-GMP then
was quantitatively converted to free [32P]phosphate
and unlabeled adenosine or guanosine by the action
15 of snake venom 5'-nucleotidase. Hence, the amount
of [32P]phosphate liberated is proportional to en-
zyme activity. The assay was performed at 30°C in a
100 µL reaction mixture containing (final concentra-
tions) 40 mM Tris HCl (pH 8.0), 1 µM ZnSO₄, 5 mM
20 MgCl₂, and 0.1 mg/mL bovine serum albumin (BSA).
Alternatively, in assays assessing PDE1-specific
activity, incubation mixtures further incorporated
the use of 0.1 mM CaCl₂ and 10 µg/mL calmodulin.
PDE enzyme was present in quantities that yield <30%
25 total hydrolysis of substrate (linear assay condi-
tions). The assay was initiated by addition of
substrate (1 mM [32P]cAMP or cGMP), and the mixture
was incubated for 12 minutes. Seventy-five (75) µg
of Crotalus atrox venom then was added, and the
30 incubation was continued for 3 minutes (15 minutes
total). The reaction was stopped by addition of 200
µL of activated charcoal (25 mg/mL suspension in 0.1
M NaH₂PO₄, pH 4). After centrifugation (750 X g for

3 minutes) to sediment the charcoal, a sample of the supernatant was taken for radioactivity determination in a scintillation counter and the PDE activity was calculated.

5 Inhibitor analyses were performed similarly to the method described in Loughney et al., *J. Biol. Chem.*, 271, pp. 796-806 (1996), except both cGMP and cAMP were used, and substrate concentrations were kept below 32 nM, which is far below the
10 Km of the tested PDEs.

Human PDE4A, 4B, 4C, 4D Preparations

Preparation of PDE4A from *S. cerevisiae*

15 Yeast cells (50 g of yeast strain Y126 harboring HDUN1.46) were thawed at room temperature by mixing with 50 mL of Lysis Buffer (50 mM MOPS pH 7.5, 10 μ M ZnSO₄, 2 mM MgCl₂, 14.2 mM 2-mercapto-
20 ethanol, 5 μ g/mL each of pepstatin, leupeptin, aprotinin, 20 μ g/mL each of calpain inhibitors I and II, and 2 mM benzamidine HCl). Cells were lysed in a French[®] pressure cell (SLM-Aminco[®], Spectronic Instruments) at 10°C. The extract was centrifuged
25 in a Beckman JA-10 rotor at 9,000 rpm for 22 minutes at 4°C. The supernatant was removed and centrifuged in a Beckman TI45 rotor at 36,000 rpm for 45 minutes at 4°C.

30 PDE4A was precipitated from the high-speed supernatant by the addition of solid ammonium sulfate (0.26 g/mL supernatant) while stirring in an ice bath and maintaining the pH between 7.0 and 7.5. The precipitated proteins containing PDE4A were

collected via centrifugation in a Beckman JA-10 rotor at 9,000 rpm for 22 minutes. The precipitate was resuspended in 50 mL of Buffer G (50 mM MOPS pH 7.5, 10 μ M ZnSO₄, 5 mM MgCl₂, 100 mM NaCl, 14.2 mM 2-mercaptoethanol, 2 mM benzamidine HCl, 5 μ g/mL each of leupeptin, pepstatin, and aprotinin, and 20 μ g/mL each of calpain inhibitors I and II) and passed through a 0.45 μ m filter.

The resuspended sample (50 to 100 mL) was loaded onto a 5 X 100 cm column of Pharmacia SEPHACRYL[®] S-300 equilibrated in Buffer G. Enzyme activity was eluted at a flow rate of 2 mL/min and pooled for later fractionation.

The PDE4A isolated from gel filtration chromatography was applied to a 1.6 X 20 cm column of Sigma Cibacron Blue Agarose-type 300 (10 mL) equilibrated in Buffer A (50 mM MOPS pH 7.5, 10 μ M ZnSO₄, 5 mM MgCl₂, 14.2 mM 2-mercaptoethanol, and 100 mM benzamidine HCl). The column was washed in succession with 50 to 100 mL of Buffer A, 20 to 30 mL of Buffer A containing 20 mM 5'-AMP, 50 to 100 mL of Buffer A containing 1.5 M NaCl, and 10 to 20 mL of Buffer C (50 mM Tris HCl pH 8, 10 μ M ZnSO₄, 14.2 mM 2-mercaptoethanol, and 2 mM benzamidine HCl). The enzyme was eluted with 20 to 30 mL of Buffer C containing 20 mM cAMP.

The PDE activity peak was pooled, and precipitated with ammonium sulfate (0.33 g/mL enzyme pool) to remove excess cyclic nucleotide. The precipitated proteins were resuspended in Buffer X (25 mM MOPS pH 7.5, 5 μ M ZnSO₄, 50 mM NaCl, 1 mM DTT, and 1 mM benzamidine HCl), and desalted via gel filtration on a Pharmacia PD-10[®] column per manufac-

turer's instructions. The enzyme was quick-frozen in a dry ice/ethanol bath and stored at -70°C.

The resultant preparations were about >80% pure by SDS-PAGE. These preparations had specific activities of about 10 to 40 μ mol cAMP hydrolyzed per minute per milligram protein.

Preparation of PDE4B from *S. cerevisiae*

10 Yeast cells (150 g of yeast strain YI23 harboring HDUN2.32) were thawed by mixing with 100 mL glass beads (0.5 mM, acid washed) and 150 mL Lysis Buffer (50 mM MOPS pH 7.2, 2 mM EDTA, 2 mM EGTA, 1 mM DTT, 2 mM benzamidinium HCl, 5 μ g/mL each
15 of pepstatin, leupeptin, aprotinin, calpain inhibitors I and II) at room temperature. The mixture was cooled to 4°C, transferred to a Bead-Beater[®], and the cells lysed by rapid mixing for 6 cycles of 30 seconds each. The homogenate was centrifuged for 22
20 minutes in a Beckman J2-21M centrifuge using a JA-10 rotor at 9,000 rpm and 4°C. The supernatant was recovered and centrifuged in a Beckman XL-80 ultracentrifuge using a TI45 rotor at 36,000 rpm for 45 minutes at 4°C. The supernatant was recovered and
25 PDE4B was precipitated by the addition of solid ammonium sulfate (0.26 g/mL supernatant) while stirring in an ice bath and maintaining the pH between 7.0 and 7.5. This mixture was then centrifuged for 22 minutes in a Beckman J2 centrifuge using a JA-10
30 rotor at 9,000 rpm (12,000 X g). The supernatant was discarded and the pellet was dissolved in 200 mL of Buffer A (50 mM MOPS pH 7.5, 5 mM MgCl₂, 1 mM DTT, 1 mM benzamidinium HCl, and 5 μ g/mL each of leupeptin,

pepstatin, and aprotinin). The pH and conductivity were corrected to 7.5 and 15-20 mS, respectively.

The resuspended sample was loaded onto a 1.6 X 200 cm column (25 mL) of Sigma Cibacron Blue Agarose-type 300 equilibrated in Buffer A. The sample was cycled through the column 4 to 6 times over the course of 12 hours. The column was washed in succession with 125 to 250 mL of Buffer A, 125 to 250 mL of Buffer A containing 1.5 M NaCl, and 25 to 50 mL of Buffer A. The enzyme was eluted with 50 to 75 mL of Buffer E (50 mM Tris HCl pH 8, 2 mM EDTA, 2 mM EGTA, 1 mM DTT, 2 mM benzamidine HCl, and 20 mM cAMP) and 50 to 75 mL of Buffer E containing 1 M NaCl. The PDE activity peak was pooled, and precipitated with ammonium sulfate (0.4 g/mL enzyme pool) to remove excess cyclic nucleotide. The precipitated proteins were resuspended in Buffer X (25 mM MOPS pH 7.5, 5 μ M ZnSO₄, 50 mM NaCl, 1 mM DTT, and 1 mM benzamidine HCl) and desalted via gel filtration on a Pharmacia PD-10[®] column per manufacturer's instructions. The enzyme pool was dialyzed overnight against Buffer X containing 50% glycerol. This enzyme was quick-frozen in a dry ice/ethanol bath and stored at -70°C.

The resultant preparations were about >90% pure by SDS-PAGE. These preparations had specific activities of about 10 to 50 μ mol cAMP hydrolyzed per minute per milligram protein.

Preparation of PDE4C from *S. cerevisiae*

Yeast cells (150 g of yeast strain Y130 harboring HDUN3.48) were thawed by mixing with 100 mL glass beads (0.5 mM, acid washed) and 150 mL Lysis Buffer (50 mM MOPS pH 7.2, 2 mM EDTA, 2 mM EGTA, 1 mM DTT, 2 mM benzamidinium HCl, 5 µg/mL each of pepstatin, leupeptin, aprotinin, calpain inhibitors I and II) at room temperature. The mixture was cooled to 4°C, transferred to a BEAD-BEATER[®], and the cells lysed by rapid mixing for 6 cycles of 30 sec each. The homogenate was centrifuged for 22 minutes in a Beckman J2-21M centrifuge using a JA-10 rotor at 9,000 rpm and 4°C. The supernatant was recovered and centrifuged in a Beckman XL-80 ultracentrifuge using a TI45 rotor at 36,000 rpm for 45 minutes at 4°C.

The supernatant was recovered and PDE4C was precipitated by the addition of solid ammonium sulfate (0.26 g/mL supernatant) while stirring in an ice bath and maintaining the pH between 7.0 and 7.5. Thirty minutes later, this mixture was centrifuged for 22 minutes in a Beckman J2 centrifuge using a JA-10 rotor at 9,000 rpm (12,000 X g). The supernatant was discarded and the pellet was dissolved in 200 mL of Buffer A (50 mM MOPS pH 7.5, 5 mM MgCl₂, 1 mM DTT, 2 mM benzamidinium HCl, and 5 µg/mL each of leupeptin, pepstatin, and aprotinin). The pH and conductivity were corrected to 7.5 and 15-20 mS, respectively.

The resuspended sample was loaded onto a 1.6 X 20 cm column (25 mL) of Sigma Cibacron Blue Agarose-type 300 equilibrated in Buffer A. The

sample was cycled through the column 4 to 6 times over the course of 12 hours. The column was washed in succession with 125 to 250 mL of Buffer A, 125 to 250 mL of Buffer A containing 1.5 M NaCl, and then 5 25 to 50 mL of Buffer A. The enzyme was eluted with 50 to 75 mL of Buffer E (50 mM Tris HCl pH 8, 2 mM EDTA, 2 mM EGTA, 1 mM DTT, 2 mM benzamidine HCl, and 20 mM cAMP) and 50 to 75 mL of Buffer E containing 1 M NaCl. The PDE4C activity peak was pooled, and 10 precipitated with ammonium sulfate (0.4 g/mL enzyme pool) to remove excess cyclic nucleotide. The precipitated proteins were resuspended in Buffer X (25 mM MOPS pH 7.2, 5 μ M ZnSO₄, 50 mM NaCl, 1 mM DTT, and 1 mM benzamidine HCl) and desalted via gel filtration on a Pharmacia PD-10[®] column per manufacturer's instructions. The enzyme pool was dialyzed 15 overnight against Buffer X containing 50% glycerol. This enzyme was quick-frozen in a dry ice/ethanol bath and stored at -70°C.

20 The resultant preparations were about >80% pure by SDS-PAGE. These preparations had specific activities of about 10 to 20 μ mol cAMP hydrolyzed per minute per milligram protein.

25 Preparation of PDE4D from *S. cerevisiae*

Yeast cells (100 g of yeast strain YI29 harboring HDUN4.11) were thawed by mixing with 150 mL glass beads (0.5 mM, acid washed) and 150 mL 30 Lysis Buffer (50 mM MOPS pH 7.2, 10 μ M ZnSO₄, 2 mM MgCl₂, 14.2 mM 2-mercaptoethanol, 2 mM benzamidine HCl, 5 μ g/mL each of pepstatin, leupeptin, aprotinin, calpain inhibitors I and II) at room tempera-

ture. The mixture was cooled to 4°C, transferred to a Bead-Beater[®], and the cells lysed by rapid mixing for 6 cycles of 30 sec each. The homogenate was centrifuged for 22 minutes in a Beckman J2-21M centrifuge using a JA-10 rotor at 9,000 rpm and 4°C. The supernatant was recovered and centrifuged in a Beckman XL-80 ultracentrifuge using a TI45 rotor at 36,000 rpm for 45 minutes at 4°C. The supernatant was recovered and PDE4D was precipitated by the addition of solid ammonium sulfate (0.33 g/mL supernatant) while stirring in an ice bath and maintaining the pH between 7.0 and 7.5. Thirty minutes later, this mixture was centrifuged for 22 minutes in a Beckman J2 centrifuge using a JA-10 rotor at 9,000 rpm (12,000 X g). The supernatant was discarded and the pellet was dissolved in 100 mL of Buffer A (50 mM MOPS pH 7.5, 10 μ M ZnSO₄, 5 mM MgCl₂, 14.2 mM 2-mercaptoethanol, 100 mM benzamidine HCl, and 5 μ g/mL each of leupeptin, pepstatin, aprotinin, calpain inhibitor I and II). The pH and conductivity were corrected to 7.5 and 15-20 mS, respectively.

At a flow rate of 0.67 mL/min, the resuspended sample was loaded onto a 1.6 X 20 cm column (10 mL) of Sigma Cibacron Blue Agarose-type 300 equilibrated in Buffer A. The column was washed in succession with 50 to 100 mL of Buffer A, 20 to 30 mL of Buffer A containing 20 mM 5'-AMP, 50 to 100 mL of Buffer A containing 1.5 M NaCl, and then 10 to 20 mL of Buffer C (50 mM Tris HCl pH 8, 10 μ M ZnSO₄, 14.2 mM 2-mercaptoethanol, 2 mM benzamidine HCl). The enzyme was eluted with 20 to 30 mL of Buffer C containing 20 mM cAMP.

The PDE4D activity peak was pooled and precipitated with ammonium sulfate (0.4 g/mL enzyme pool) to remove excess cyclic nucleotide. The precipitated proteins were resuspended in Buffer X (25 mM MOPS pH 7.2, 5 μ M ZnSO₄, 50 mM NaCl, 1 mM DTT, and 1 mM benzamidine HCl) and desalted via gel filtration on a Pharmacia PD-10[®] column per manufacturer's instructions. The enzyme pool was dialyzed overnight against Buffer X containing 50% glycerol. This enzyme preparation was quick-frozen in a dry ice/ethanol bath and stored at -70°C.

The resultant preparations were about >80% pure by SDS-PAGE. These preparations had specific activities of about 20 to 50 μ mol cAMP hydrolyzed per minute per milligram protein.

Lipopolysaccharide-Stimulated TNF α Release from Human Peripheral Blood Lymphocytes

To assess the ability of a compound to reduce TNF α secretion in human peripheral blood lymphocytes (PBL), the following tests were performed. Previous studies have demonstrated that incubation of human PBL with cAMP-elevating agents, such as prostaglandin E₂, forskolin, 8-bromo-cAMP, or dibutyl-cAMP, inhibits the secretion of TNF α by the cells when stimulated by lipopolysaccharide (LPS; endotoxin). Accordingly, preliminary experiments have been performed to demonstrate that selective PDE4 inhibitors, such as rolipram, inhibit LPS-induced TNF α secretion from human lymphocytes in a dose-dependent fashion. Hence, TNF α secretion from human PBL was used as a standard for the ability of

a compound to elevate intracellular cAMP concentrations and/or to inhibit PDE4 activity within the cell.

Heparinized blood (approximately 30 mL) drawn from human volunteers was mixed 1:1 with Dulbecco's modified phosphate-buffered saline. This mixture was mixed 1:1 with HISTOPAQUE[®] and centrifuged at 1,500 rpm at room temperature without braking in the swinging bucket of a Beckman model TJ6 centrifuge. Erythrocytes were centrifuged to the bottom of the tubes, and serum remained at the surface of the tubes. A layer containing lymphocytes sedimented between the serum and HISTOPAQUE[®] layers, and was removed by aspiration to a fresh tube. The cells were quantified and adjusted to 3×10^6 cells/mL and a 100 μ L aliquot is placed into the wells of a 96 well plate. Test compounds and RPMI media (Gibco/BRL Life Sciences) are added to each of the wells 15 minutes prior to addition of bacterial LPS (25 mg/mL). The mixture was allowed to incubate for 20 hours at 37°C in a humidified chamber. The cells then were separated by centrifuging at 800 rpm for 5 minutes at room temperature. An aliquot of 180 μ L of supernatant was transferred to a new plate for determination of TNF α concentration. TNF α protein in the cell supernatant fluids was measured using a commercially available enzyme-linked immunosorbent assay (ELISA) (CYTOSCREEN[®] Immunoassay Kit from Biosource International).

The cell-based assay provided the following results for various pyrrolidine compounds of the present invention. The EC₅₀ values (i.e., effective concentration of the compound capable of inhibiting

50% of the total TNF α) illustrate the ability of the present compounds to inhibit LPS-stimulated TNF α release from human PBL.

The table below illustrates the ability of
5 compounds of formula (II) to inhibit PDE4 activity and TNF α release *in vitro*. In the following table, the IC₅₀ values were determined against human recombinant PDE4..

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	Example Number ¹⁾	PDE4 IC ₅₀ (M x 10 ⁻⁹)	PBL/TNFα EC ₅₀ (M x 10 ⁻⁹)
	1	1400.0	775.5
	2	28.5	142.0
5	3	3134.1	
	4	3091.7	
	5	2513.9	
	6	506.5	730.0
	7	57.0	52.5
10	8	673.2	1562.3
	9	57.5	608.7
	10	2951.1	
	11	27,050.7	
	12	14,695.9	
15	13	2435.0	
	14	529.2	598.2
	15	8967.0	
	16	9075.7	
	17	8663.4	
20	18	7392.7	
	19	51.7	59.0
	20	22.0	198.3
	21	2204.7	
	22	5888.2	
25	23	7549.3	
	24	22.9	76.5
	25	53.0	85.5
	26	235.1	450.0
	27	479.3	816.0
30	28	590.9	1392.0

The data presented above shows that the present compounds are potent inhibitors of PDE4, e.g., the compounds have an IC_{50} vs. human recombinant PDE4 of about 700 pM to about 15 μ M. Preferred compounds have an IC_{50} of about 100 nM or less, and especially preferred compounds have an IC_{50} of about 50 nM or less.

Similarly, preferred compounds have a PBL/TNF α EC_{50} about 500 nM or less, and preferably about 200 nM or less. More preferred compounds have a PBL/TNF α of about 100 nM or less.

To achieve the full advantage of the present invention, the compound has an IC_{50} vs. human recombinant PDE4 of about 100 μ M or less and a PBL/TNF α EC_{50} of about 500 μ M or less. More preferably, the compound has an IC_{50} of about 50 nM or less and a PBL/TNF α EC_{50} of about 100 nM or less.

Animal Models

20 Combined Mouse endotoxin-stimulated TNF α Release and Locomotor Activity Assay

The purpose of this study was to determine the efficacy of PDE4 inhibitors *in vivo* in an LPS mouse model together with a determination with respect to central nervous system (CNS) side-effects manifested by a decrease in spontaneous mobility.

The test animals were female Balb/c mice, having an average weight of about 20 g. The PDE4 inhibitors, formulated in 30% Cremophor[®] EL, were administered via intraperitoneal (i.p.) injections at doses of 0.1, 1.0, 10.0, and 100 mg/kg. Individual dose volumes (about 150 μ L) were adjusted based

on the body weights measured. One hour later, 5
mg/kg LPS in a final volume of 200 μ L was injected
via the tail vein to each animal. Ninety minutes
following the LPS treatment, the animals were bled
5 and serum samples were collected before being stored
at -70°C until assayed.

For efficacy determination, the serum
samples were diluted two-fold and TNF α levels were
determined using the CYTOSCREEN[®] Immunoassay Kit
10 (Biosource International). The data were averaged
between triplicate sample subjects for each of the
tested compounds.

Movement of the X-Y plane, or rearing up
on the hind legs, was quantified by counting the
15 number of "light-beam" crosses per unit of time. A
decrease in the number of activity events is di-
rectly proportional to the mobility or immobiliza-
tion of the animal. The quantitative scoring corre-
lated well with the subjective measurements de-
20 scribed above.

The following table summarizes the mouse
locomotor activity assay as mobility (% activity)
obtained by the above-described method:

25	Example Number¹⁾	Mobility (% activity at 50 mg/kg dose)
	7	16%
	9	55%
	19	145%
30	20	93%

It also was determined that compounds of
35 formula (II) have fewer central nervous system side

effects compared to rolipram and to compounds disclosed in Feldman et al. U.S. Patent No. 5,665,754. It also was found that central nervous system activity is related to the absolute stereochemistry of
5 the present compounds.

The results summarized above show that the compounds of the present invention are useful for selectively inhibiting PDE4 activity in a mammal, without exhibiting the adverse CNS and emetic effects associated with prior PDE4 inhibitors.
10

Obviously, many modifications and variations of the invention as hereinbefore set forth can be made without departing from the spirit and scope thereof and, therefore, only such limitations should
15 be imposed as are indicated by the appended claims.

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